

ORIGINAL ARTICLE

Signals of differential introgression in the genome of natural hybrids of Caribbean anoles

Keaka Farleigh¹  | Alfredo Ascanio¹  | Mackenzie E. Farleigh¹  | Drew R. Schield²  |
Daren C. Card^{3,4}  | Manuel Leal⁵ | Todd A. Castoe⁶  | Tereza Jezkova¹  |
Javier A. Rodríguez-Robles⁷ 

¹Department of Biology, Miami University, Oxford, Ohio, USA

²Department of Ecology and Evolutionary Biology, University of Colorado, Boulder, Colorado, USA

³Department of Organismic and Evolutionary Biology, Harvard University, Cambridge, Massachusetts, USA

⁴Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, USA

⁵Division of Biological Sciences, University of Missouri, Columbia, Missouri, USA

⁶Department of Biology, University of Texas, Arlington, Arlington, Texas, USA

⁷School of Life Sciences, University of Nevada, Las Vegas, Las Vegas, Nevada, USA

Correspondence

Keaka Farleigh, Department of Biology, Miami University, Oxford, OH 45056 USA.
Email: farleik@miamioh.edu

Funding information

Miami University; National Science Foundation, Grant/Award Number: DEB#x2010;1655571 and IOS#x2010;1051793; National Science Foundation Graduate Research Fellowship Program, Grant/Award Number: 2037786; University of Nevada, Las Vegas; University of Texas, Arlington

Handling Editor: Nick Hamilton Barton

Abstract

Hybridization facilitates recombination between divergent genetic lineages and can be shaped by both neutral and selective processes. Upon hybridization, loci with no net fitness effects introgress randomly from parental species into the genomes of hybrid individuals. Conversely, alleles from one parental species at some loci may provide a selective advantage to hybrids, resulting in patterns of introgression that do not conform to random expectations. We investigated genomic patterns of differential introgression in natural hybrids of two species of Caribbean anoles, *Anolis pulchellus* and *A. krugi* in Puerto Rico. Hybrids exhibit *A. pulchellus* phenotypes but possess *A. krugi* mitochondrial DNA, originated from multiple, independent hybridization events, and appear to have replaced pure *A. pulchellus* across a large area in western Puerto Rico. Combining genome-wide SNP datasets with bioinformatic methods to identify signals of differential introgression in hybrids, we demonstrate that the genomes of hybrids are dominated by *pulchellus*-derived alleles and show only 10%–20% *A. krugi* ancestry. The majority of *A. krugi* loci in hybrids exhibit a signal of non-random differential introgression and include loci linked to genes involved in development and immune function. Three of these genes (delta like canonical notch ligand 1, jagged1 and notch receptor 1) affect cell differentiation and growth and interact with mitochondrial function. Our results suggest that differential non-random introgression for a subset of loci may be driven by selection favouring the inheritance of compatible mitochondrial and nuclear-encoded genes in hybrids.

KEYWORDS

Anolis, *HybridFindR*, hybridization, introgression, single nucleotide polymorphisms, West Indies

1 | INTRODUCTION

Hybridization is relatively common between closely related species that lack complete reproductive isolation (Capblancq et al., 2019; Mallet, 2005; Nosil, 2012). Hybridization enables recombination between lineages that come into contact, which can lead to genetic homogenization and impede or reverse the process of divergence (Abbott et al., 2013; Gow et al., 2006; Seehausen, 2006; Taylor et al., 2006). Hybrids often exhibit lower fitness than parental species due to endogenous selection (e.g., genetic incompatibilities between parental genomes) or exogenous selection (e.g., inferior fitness in local environmental conditions), processes that can limit hybrid persistence (Barton, 2001; Fierst & Hansen, 2010; Seehausen, 2004). However, in some cases, hybrids may exhibit higher fitness than parental species because of the acquisition of functional novelty or a new combination of advantageous traits (Seehausen, 2004). These novel features can provide fitness advantages to hybrids, for example, by optimizing cellular processes such as metabolism and immune response or by facilitating adaptation to distinct thermal environments (Martin et al., 2020). While hybrids typically express phenotypes that are combinations of parental traits (Grant & Grant, 1994), in some cases, hybrids may instead exhibit mostly or entirely the phenotype of a single parental species (Ferris et al., 1983; Hird & Sullivan, 2009; Jezkova et al., 2013a; Liu et al., 2010; Tegelström, 1987). The absence of obvious phenotypic introgression in hybrids has generally been attributed to either extensive directional backcrossing (i.e., breeding of the hybrids with only one parental species) or reduced introgression between hybridizing taxa due to major chromosomal incompatibilities (Llopart et al., 2005; Rieseberg et al., 1996).

Genetic introgression describes the incorporation of alleles from one population into the gene pool of another population (Anderson, 1949; Anderson & Hubricht, 1938; Harrison & Larson, 2014). Herein, we use the terms “hybridization” and “genetic introgression” to describe separate but related phenomena: hybridization refers to the process of interspecific mating, whereas genetic introgression describes the genetic outcome of hybridization and subsequent backcrossing and natural selection altering the frequencies of subsets of hybrid genotypes. During introgression, alleles that are not targeted by selection or that do not have incompatibilities with the recipient genomic background are expected to introgress freely, assuming that recombination occurs between these and the physically linked targets of selection. We refer to these loci as exhibiting “neutral introgression” (Figure 1), where a hybrid is equally likely to inherit an allele from either parental population. In this scenario, the only expected directional pattern is a decrease in the number of introgressed alleles from one parental population as the frequency of backcrossing with the second parental population increases.

Alternatively, if alleles (or allele combinations across multiple loci) from one parental species increase the relative fitness of individuals in the other parental population, these alleles are more likely to introgress (Harrison & Larson, 2014; Hedrick, 2013;

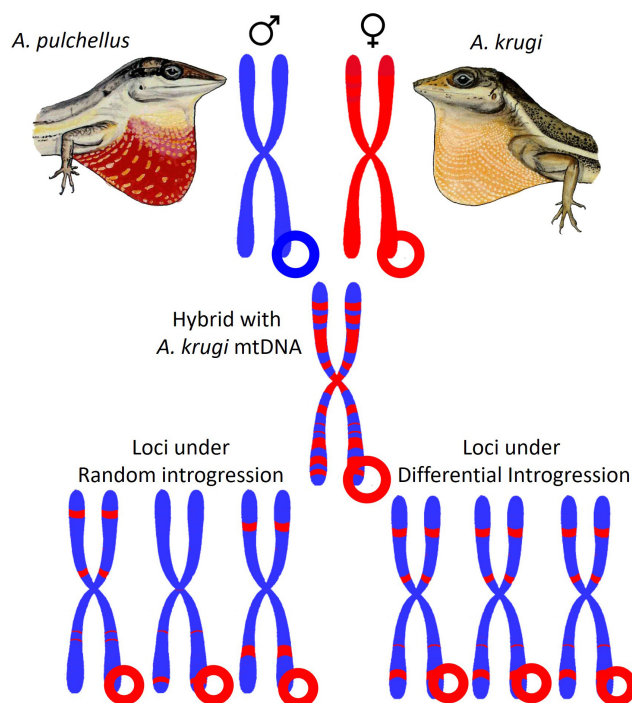


FIGURE 1 Visualization of neutral introgression and differential introgression scenarios with respect to the extent and pattern of nuclear introgression in *Anolis pulchellus* hybrids (*A. pulchellus* with *A. krugi* mtDNA). In neutral introgression, matings between *A. pulchellus* males and *A. krugi* females produce hybrids that exhibit genetic introgression. When alleles are not under selection, introgression occurs indiscriminately across the genome, leading to a pattern of random introgression among hybrids. In this scenario, the introgressed *A. krugi* alleles will differ among hybrids. In differential introgression, loci are under selection (or linked to a gene or genes under selection), leading to non-random introgression that manifests as identical or enriched patterns of alleles from *A. krugi* in the hybrids. Blue = *A. pulchellus* markers; red = *A. krugi* markers. Blue circles = *A. pulchellus* mtDNA; red circles = *A. krugi* mtDNA.

Morales et al., 2017; Pardo-Diaz et al., 2012; Whitney et al., 2006); we refer to these loci as exhibiting “differential introgression” (e.g., Payseur, 2010; Walsh et al., 2016; Figure 1). Differentially introgressed loci may contain or be linked to alleles that confer a selective advantage to the hybrids in their local environment (Hedrick, 2013). These loci may also show patterns of differential introgression because they are involved in complex co-evolved gene interactions (Pardo-Diaz et al., 2012, 2015). For example, mitochondrial-nuclear interactions are fundamental to individual fitness and various biological processes (Burton, 2022; Burton & Barreto, 2012; Chang et al., 2016), including energy production and cellular respiration, both of which depend on coordinated operation between nuclear and mitochondrial processes, a coordination that can be disrupted by introgression (Wolff et al., 2014). As a result, selection will act to optimize these interactions (i.e., positive selection) and eliminate deleterious combinations of interacting alleles (i.e., purifying selection; Hill et al., 2019; Wolff et al., 2014). It is possible to detect differential introgression by searching for

alleles from one species that are at a higher frequency in hybrids than expected at random (Ballard & Melvin, 2010; Boratyński et al., 2014; Doiron et al., 2002; Llopart et al., 2005). In practice, finding conclusive evidence of differential introgression is challenging, especially in natural populations.

Anolis lizards (anoles) in the Caribbean islands are one of the best-known examples of adaptive radiation, with more than 150 currently recognized species (Losos, 2009; Losos et al., 1998; Schluter, 2000). Caribbean anole communities can be highly diverse, with up to 11 species coexisting in a single locality (Warheit et al., 1999). Despite frequent interspecific interactions, hybridization between anole species seems to be rare, and apparent hybridization has only been suggested in eight species pairs (Losos, 2009). The rarity of interspecific hybridization in anoles has been attributed to strong premating isolation mechanisms mediated by effective visual species recognition signals that prevent interspecific mating (Leal & Losos, 2015; Losos, 2009).

One such rare example of interspecific hybridization in anoles occurs between the sister species *Anolis pulchellus* (Puerto Rican Bush Anole) and *A. krugi* (Olive Bush Anole) in Puerto Rico (Jezkova et al., 2013a). These two species are often syntopic across Puerto Rico and are distinguished by their distinctive body-colour patterns and the coloration of their dewlaps (Figure 1), extensible throat fans used for intra- and interspecific communication (Fleishman et al., 2009, 2016; Leal, 1999). Both species are locally abundant and widespread; *A. pulchellus* generally occurs in grassy, sunny, warmer habitats, whereas *A. krugi* is typically associated with forested or forest-edge, partially shaded, cooler areas (Fleishman et al., 2009; Gunderson et al., 2018; Rivero, 1998; Schwartz & Henderson, 1991; Williams, 1983). A previous study documented unidirectional mtDNA introgression between these two species and inferred that hybridization between these anoles had occurred recently and independently at numerous localities (a minimum of 10 independent hybridization events were estimated; Jezkova et al., 2013a). Individuals of *A. pulchellus* with foreign (*A. krugi*) mtDNA phenotypically resemble "pure" *A. pulchellus* (those with native mtDNA, hereafter referred to simply as "*A. pulchellus*"), and large parts of western Puerto Rico are exclusively inhabited by *A. pulchellus* harbouring *A. krugi* mtDNA, suggesting that the latter may have a selective advantage over pure *A. pulchellus* (Jezkova et al., 2013a). However, the nature of the putative selective advantage of *A. krugi* mtDNA or patterns of nuclear genomic variation in these hybrids has not been explored.

In this study, we used genome-wide sampling of single nucleotide polymorphisms (SNPs) to investigate the nuclear genomic composition of *A. pulchellus* that carries foreign (*A. krugi*) mtDNA (hereafter referred to as "hybrids"). We evaluated the extent to which these hybrids possess nuclear alleles from each parental species and gathered evidence for neutral and differential introgression of *A. krugi* alleles into the nuclear genome of the hybrids. Alleles experiencing neutral or differential introgression are predicted to yield distinctive genetic signatures (Figure 1). Alleles exhibiting neutral introgression are expected to be present in a random subset of the hybrids, due to stochasticity and a varying number of backcrossing events since hybridization (Figure 1). In contrast, alleles with evidence of differential introgression should

be present in the majority of the hybrids (Llopart et al., 2005, 2014; Rieseberg, 2011; Rieseberg et al., 1996). We identified SNPs with signatures of differential introgression using a methodology that we developed and made available in the R package *HybridFindR*. We also identified signatures of differential introgression using existing methods that account for the demographic histories of parental and hybrid populations. We identified genes linked to each SNP that exhibited a signal of differential introgression by mapping our data to the genome of the congeneric species *A. carolinensis* (Alföldi et al., 2011). Finally, we conducted gene ontology (GO) analysis to identify molecular functions associated with those genes. We hypothesized that the introgression of *A. krugi* alleles into the nuclear genome of the hybrids is non-neutral and that a subset of introgressed alleles exhibits signals of differential introgression and is associated with genes that influence mitochondrial function (Jezkova et al., 2013a).

2 | MATERIALS AND METHODS

2.1 | Sampling design

We generated two SNP datasets, a medium-density (MD) dataset targeting ~10,000 loci/individual for 80 individuals and a high-density (HD) dataset targeting ~150,000 loci/individual for 10 individuals (Figure 2, Table S1), to complement mtDNA datasets previously developed for 211 *A. krugi* (Rodríguez-Robles et al., 2010) and 309 *A. pulchellus* and hybrids (Jezkova et al., 2013a). For the MD SNP dataset, samples included hybrids ($n=24$ samples, 9 localities representing ≥ 10 independent hybridization events), *A. pulchellus* ($n=27$, 10 localities) and *A. krugi* ($n=29$, 10 localities; Figure 2 and Table S1). This sampling encompasses the major mitochondrial lineages, spans the geographic distribution of each group (*A. pulchellus*, *A. krugi*, hybrids), and includes individuals with published mtDNA sequences (Figure 2; Jezkova et al., 2013a; Rodríguez-Robles et al., 2010). The HD dataset consisted of four hybrids representing four independent introgression events: two *A. pulchellus* and four *A. krugi* (Figure 2 and Table S1). We assumed that hybrids between *A. pulchellus* and *A. krugi* can be determined using mtDNA sequences and that wherever hybridization between *A. pulchellus* and *A. krugi* occurred, *A. pulchellus* harbours *A. krugi* mtDNA haplotypes. Hybrids and *A. pulchellus* are geographically separated, with populations of hybrids being restricted to the western part of Puerto Rico and *A. pulchellus* occurring throughout the remainder of the island. Nevertheless, hybrid and pure *A. pulchellus* are known to co-occur at one locality in central western Puerto Rico (locality 17; Figure 2).

2.2 | Laboratory methods

We generated the MD genome-wide SNP dataset using double-digest RAD (restriction-site associated DNA) sequencing (ddRAD-seq; Jones et al., 2012; Peterson et al., 2012). Genomic DNA was simultaneously cut with the rare-cutting restriction enzyme *SbfI*

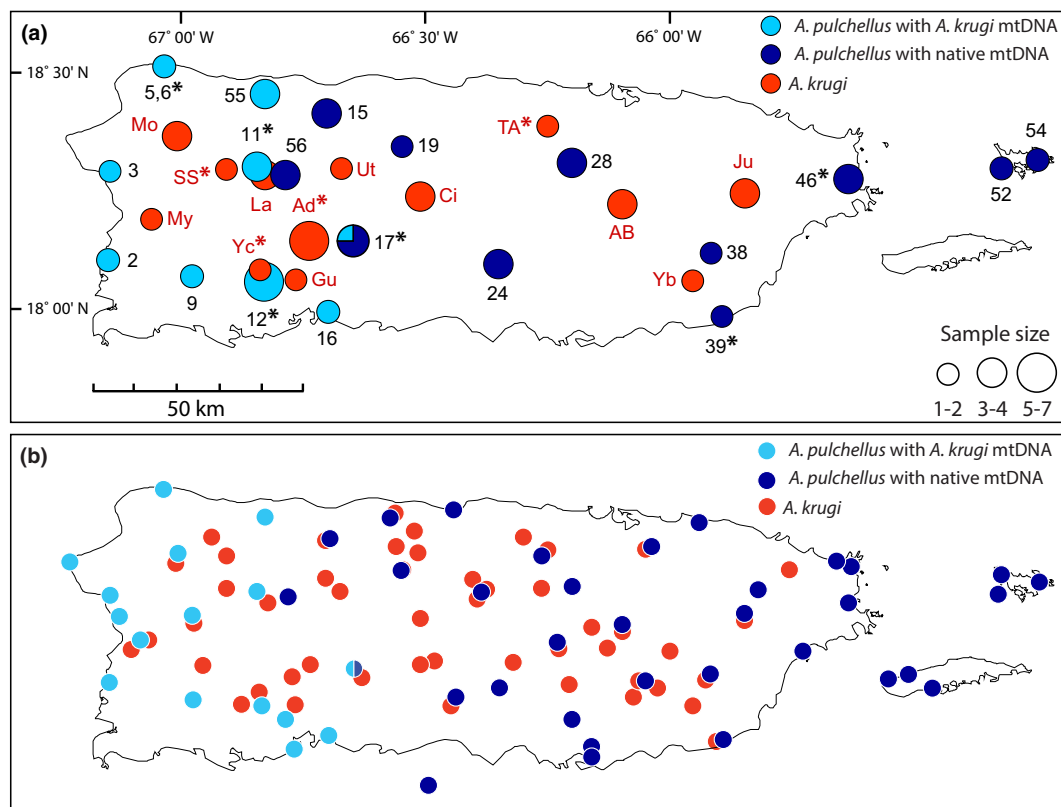


FIGURE 2 (a) Approximate sampling localities of individuals used to generate the medium-density ($N=73$) and high-density ($N=10$) SNP datasets used in this study: *Anolis pulchellus* (*A. pulchellus* with native mtDNA, dark blue circles, $N=25$), hybrids (*A. pulchellus* with *A. krugi* mtDNA, light blue circles, $N=23$) and *A. krugi* (red circles, $N=25$). Circle size is proportional to sample size. The localities for which a single sample was used for the high-density SNP dataset ($N=10$) are indicated with an asterisk (*) next to the locality code. The locality codes of *A. pulchellus* are numerical, whereas the locality codes of *A. krugi* represent municipality abbreviations, following Jezkova et al. (2013a) and Rodríguez-Robles et al. (2010). The list of all samples and sampling localities is listed in Table S1. (b) Approximate sampling localities with known mtDNA assignments for *A. pulchellus* (dark blue circles), *A. krugi* (red circles) and hybrids (light blue circles) identified in Jezkova et al. (2013a).

(5'-CCTGCA*G+G-3'; New England Biolabs Inc.) and the common-cutting restriction enzyme *Sau3AI* (5'-*GATC -3'; New England Biolabs Inc.). Barcoded Illumina adapter oligonucleotides, which included an 8 bp unique molecular identifier, were ligated to the ends of digested DNA to allow for hierarchical pooling, multiplexing of samples and PCR clone filtering. Following adapter ligation, samples were pooled into sets of eight and size-selected for a range of 340–600 bp using a Blue Pippin device (Sage Science). After size selection, samples were amplified with pool-specific indexed primers and amplification products were further pooled into two groups (based on molarity calculations) on a Bioanalyzer (Agilent) using a DNA 7500 chip. We sequenced the final pooled libraries using 100 bp paired-end reads on an Illumina HiSeq 2000 lane.

We obtained the HD SNP data using single-digest RAD sequences from Floragenex, Oregon, which generated and sequenced RAD libraries following the methods outlined by Baird et al. (2008), Hohenlohe et al. (2010) and Emerson et al. (2010). Genomic DNA was cut with the *PstI* restriction enzyme; adaptor oligonucleotides and individual barcodes were ligated to the ends of digested DNA; and the resulting fragments were sequenced on the Illumina GAIIx platform with single-end 80-bp chemistry.

For both RAD datasets, we removed PCR clones, quality-filtered and demultiplexed read data using the functions *clone_filter* and *process_radtags* in Stacks v1.42 (Catchen et al., 2013), requiring a minimum Phred quality score of 33, and using default parameters for all other options. For both MD and HD datasets, we used the dDocent pipeline for quality trimming and for mapping the reads to the assembled genome of *A. carolinensis* (genome assembly AnoCar2.0; 1,78 Gb; Alföldi et al., 2011) using BWA (Li, 2013; see Table S1 for information on genome assembly). We filtered the variants produced by dDocent using FreeBayes (Garrison & Marth, 2012), following the general steps recommended in the dDocent documentation (Puritz, Hollenbeck, et al., 2014; Puritz, Matz, et al., 2014) using vcftools and vcflib (Danecek et al., 2011). We also filtered variants (i.e., single-nucleotide polymorphisms; SNPs) to include those called in 50% of individuals ($--max-missing\ 0.50$) and with a minimum mean read depth among samples of 5 ($--min-meanDP\ 5$). Individuals with 60% or greater missing data were excluded, and we removed loci with minor allele frequencies lower than 0.05 ($--maf\ 0.05$) and SNPs that were in strong linkage disequilibrium (LD; $Rho \geq 0.96$). LD was estimated using a custom R script developed by Finger et al. (2022). When SNPs were found to be in LD, we randomly selected one to include

in our analysis. We also analysed the sequences in dDocent de novo, that is, without mapping them first to the *A. carolinensis* reference genome (Alföldi et al., 2011). The de novo approach allowed us to analyse reads that could not be mapped due to the divergence between *A. carolinensis* and *A. krugi* and *A. puchellus* (see Supplemental Information for detailed de novo methods and results).

2.3 | Analyses of the SNP datasets

We conducted a principal component analysis (PCA) separately on each dataset (the MD and HD SNP datasets) using the R package *adeigenet* v.2.1.5 (Jombart, 2008) available in R v4.0.1 (R Core Team, 2020). We also constructed a Neighbour-Net (Bryant & Moulton, 2004) for the MD dataset and for the mtDNA for the 73 individuals included in the MD dataset using the R package *phangorn* v.2.8.0 (Schliep, 2011) available in R v4.0.1 (R Core Team, 2020). We downloaded the mtDNA sequences from the Dryad digital repository (<https://doi.org/10.5061/dryad.304dn>), developed by Jezkova et al. (2013b).

We estimated the individual hybrid index (HI) and 95% confidence intervals for each dataset using bi-allelic SNPs and the Bayesian Markov chain Monte Carlo (MCMC) method implemented in the R-package *gghybrid* v.1.0.0 (Bailey, 2020). The HI represents an estimate of the proportion of alleles that were inherited from one of

the two parental species and is dependent on the allele frequencies of parental populations (Buerkle, 2005). HI estimates range from 0 to 1, with extreme values (i.e., values close to 0 or 1) corresponding to pure individuals of each parental species. When parental individuals are fixed for different alleles and half of the hybrid alleles are from either parental population, we expect the HI to be 0.5 for all hybrids (see Table S2 for a visualization). Alternatively, when parental populations (S0 and S1 populations; S = Source) are polymorphic and the S0 population possesses S1 alleles, HINDEX estimates are no longer 0.5, even though hybrid genotypes are half S0 and half S1 alleles (see Table S2 for a visualization). We used the recommended settings and filtered our datasets using a minimum minor allele frequency of 0.1 and a minimum allele frequency difference (minimum difference between parental populations) of 0.25 to remove uninformative loci. We ran 10,000 MCMC iterations with a burn-in of 5000 iterations and visually inspected the MCMC chains to ensure mixing and convergence (Figure S1). We assigned localities of *A. krugi* and *A. puchellus* with native mtDNA as parental populations. Parental limits are calculated using the 95% confidence intervals of the parental populations and are defined as the maximum value for the S0 population (*A. pulchellus*) and the minimum value for the S1 population (*A. krugi*). These limits are used to determine whether hybrid individuals are more similar to one source population (i.e., whether the hybrids fall within the parental limits) or whether they are a mix of the two parental populations (i.e., hybrids fall outside the parental

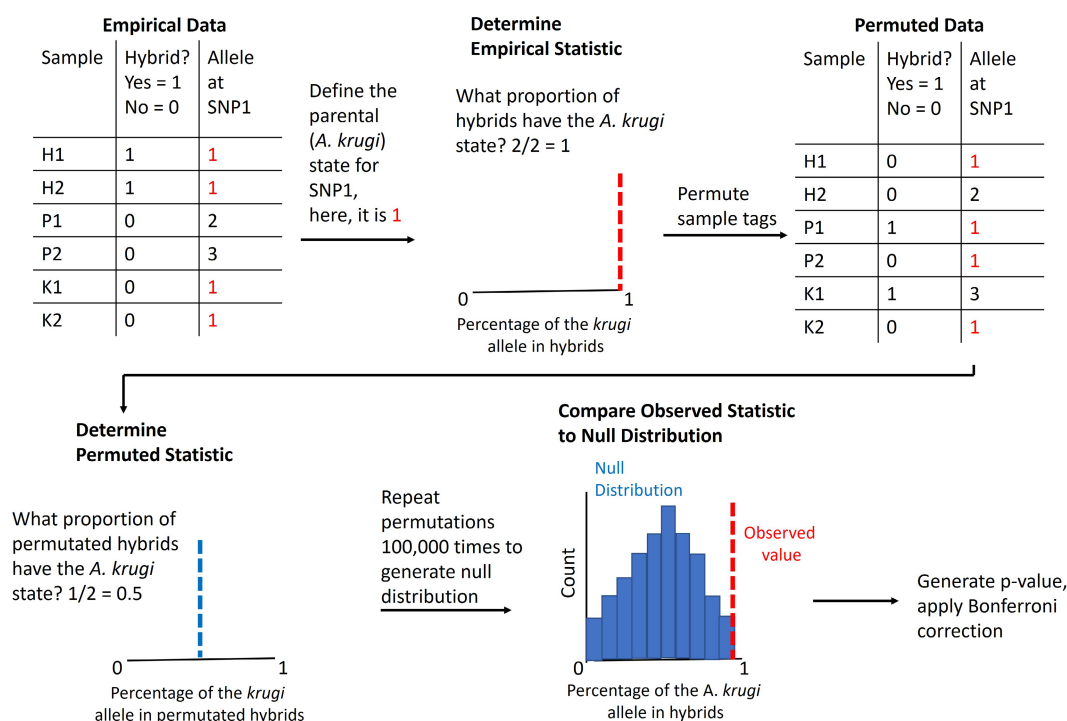


FIGURE 3 Visualization of the procedure used in *HybridFindR* to identify SNPs exhibiting signals of differential introgression in *Anolis pulchellus* hybrids (*A. pulchellus* with *A. krugi* mtDNA). First, we used *gghybrid* (Bailey, 2020) to identify the *A. krugi* allele for each SNP. We then determined the proportion of *A. krugi* alleles in the hybrids to calculate the observed statistic. Next, we permuted the population tags and estimated the proportion of *A. krugi* alleles in permuted hybrids to calculate the permuted statistic. Permutations were repeated 100,000 times to generate a null distribution. Finally, we compared the observed statistic to the null distribution, calculated a *p*-value, and applied the Bonferroni correction to identify SNPs that may exhibit signals of differential introgression.

limits). This method accommodates codominant markers and markers that are not fixed between taxa, making it appropriate for closely related species (Buerkle, 2005). We first estimated the HI for the entire SNP dataset and then separately for each chromosome to determine whether the HI estimation was consistent across the nuclear genome. Due to the small number of SNPs on individual microchromosomes, we considered all SNPs on microchromosomes as a single group for comparative purposes.

We developed a new approach to test whether the observed pattern of differential introgression events across multiple individuals was likely due to random chance or if instead selection may be promoting the differential introgression of individual SNPs (Figure 3). This approach was developed because hybrids do not form a typical clinal hybrid zone between parapatrically distributed taxa, where the degree of introgression decreases with the distance from the contact zone. Therefore, our dataset is not suitable for geographic cline analysis that requires a wide range of HI estimates along the hybrid zone. Our procedure tests the null hypothesis that at each SNP, the proportion of alleles from one parental species in the genomes of the hybrids is due to neutral introgression. This assessment is achieved by using permutations of population labels of individuals (hybrids vs. parental); it calculates the proportions of the selected set of parental alleles in permuted hybrids and then compares these proportions with the empirical proportions observed from the selected parental alleles in the hybrid samples. We assigned *A. krugi* as the parental species to perform our comparisons, and for each hybrid SNP we identified the corresponding parental allele using source alleles (alleles from parental populations) previously determined by HI estimation. For each hybrid SNP, we calculated the proportion of *A. krugi* alleles in hybrids to represent our observed statistic. Next, we permuted the population labels of individuals 100,000 times and calculated the proportion of *A. krugi* alleles in permuted hybrids, to generate a null distribution of permutations' proportions. We then calculated a p-value as the proportion of random permutations that yielded a frequency of introgressed alleles greater than the frequency derived from the observed data. We corrected our data for multiple comparisons using the Bonferroni method (Dunn, 1961) and considered any SNP with a corrected p-value less than 0.05 to be a candidate SNP exhibiting differential introgression. We then calculated the percentage of *A. krugi* and *A. pulchellus* alleles that were randomly and differentially introgressed in the genomes of the hybrids. For example, if 367 of 3140 SNPs were differentially introgressed with an upper HI index of 0.161 (16.1% of *A. krugi* loci in the hybrid genome), then 11.6% (367/3140) of *A. krugi* alleles were differentially introgressed and 4.5% (16.1%–11.6%) were randomly introgressed. We compiled these methods into the R-package *HybridFindR*, which is available on GitHub (<https://github.com/kfarleigh/HybridFindR>).

One drawback of our analysis is that it does not account for the demographic histories of parental and hybrid populations. Therefore, we assessed whether genetic drift could account for the observed distribution of the introgressed alleles. We did this in two steps. First, we performed neutral demographic simulations to test competing hypotheses for patterns of diversification

among *A. pulchellus*, *A. krugi* and hybrid populations using three-dimensional (3D) joint frequency spectra (JSFS) of genetic variation using the program *moments* v.1.1.13 (Jouanous et al., 2017). We tested five models that differ in the patterns of gene flow (asymmetric or symmetric gene flow; Figure S2) between parental and hybrid populations. We built folded JSFS and down-sampled genotypes to minimize missing data and maximize the number of segregating sites using the program *easySFS* (<https://github.com/isaacovercast/easySFS>). Models were optimized and ran following the methods of Leaché et al. (2019), with python scripts for performing model fitting downloaded from github.com/dportik/moments_pipeline (Portik et al., 2017). We performed modelling with folded JSFS because outgroup data were not available. We conducted four rounds of optimizations for each model, using the parameters from the best replicate as starting values for the next round of optimization. Replicate analyses were carried out to ensure that the optimization routine was stable. Before the final analysis, we used the replicate with the highest likelihood for each model to calculate Akaike information criterion (AIC) scores (Burnham & Anderson, 2004), and the model with the lowest AIC was considered the best model. We then converted parameter estimates of theta (θ) and migration (M_{ij}) for the best model to effective population sizes (N_e) using the equation $N_e = \theta/4\mu L$, where μ represents the mutation rate and L is the number of loci multiplied by their length. We estimated migration rates between populations (m_{ij}) with the equation $m_{ij} = M_{ij}/2N_e$ and divergence times (t) using the equation $t = T/2N_e g$, where g is generation time. We used the *A. carolinensis* mutation rate (2.1×10^{-10} ; Bourgeois et al., 2019) and a generation time of 1 year in our calculations (Tollis & Boissinot, 2014).

In the second step, we used coalescent simulations to determine whether the observed allele frequencies could be generated by genetic drift. We used parameters inferred by the best-performing model in the previous step as input into *msprime* v.1.0.2 (Baumdicker et al., 2022). We simulated 1000,000 loci across two separate runs for each dataset. Simulations were processed with *PGDspider* v.2.1.1.5 (Lischer & Excoffier, 2012); custom scripts are available at <https://github.com/kfarleigh/AnolisSims>. We used processed files from each simulation run as input into *gghybrid*, calculated the observed proportion of parental alleles in the hybrids and compared the distribution of simulated frequencies to our empirical distribution. We compared distributions with the Kolmogorov–Smirnov test (Massey, 1951) to determine whether the distributions were significantly different. We compared the two simulated distributions to ensure that they yielded consistent results.

A second potential drawback of our approach is that the null distribution generated by permutations can be influenced by the sample sizes of the populations and the amount of backcrossing. Nevertheless, our sample sizes were approximately equal (*A. krugi*, 25; *A. pulchellus*, 25; Hybrids, 23), and because the levels of backcrossing in the hybrids are unknown, we did not include this parameter in our model, which should result in a more conservative test of differential introgression.

We identified loci harbouring SNPs with a signal of differential introgression and linked those loci to known genes and their associated functions based on the *A. carolinensis* genome annotation (Alföldi et al., 2011). We used the Genome Data Viewer (https://www.ncbi.nlm.nih.gov/genome/gdv/browser/genome/?id=GCF_000090745.1) to determine whether an SNP falls within an annotated gene transcript in *A. carolinensis*. In instances where an SNP occurred within an annotated gene transcript, we performed GO analyses of genes containing these SNPs using QuickGO (Binns et al., 2009) to infer their possible biological relevance to hybrids (see Appendix S1 for methods used to annotate de novo candidate SNPs exhibiting differential introgression).

3 | RESULTS

3.1 | ddRADseq and filtering

We generated two SNP datasets: a medium-density dataset (MD) for 80 individuals using ddRADseq (Jones et al., 2012; Peterson et al., 2012) and a high-density (HD) dataset for 10 individuals using single-end RAD sequencing (Figure 2, Table S1; Baird et al., 2008). We then created two datasets each for the MD and HD datasets by mapping sequence reads onto the genome of *A. carolinensis* (reference-based; Alföldi et al., 2011) and by clustering reads de novo (i.e., reference-independent; see Section 2.2 for details). After removing individuals with greater than 60% missing data, we were left with 73 individuals in the MD dataset and 10 individuals in the HD dataset (Figure 2, Table S1). The datasets comprised 3140 SNPs and 14,444 SNPs when reads were mapped to the *A. carolinensis* genome (Alföldi et al., 2011), with an average depth of 7.5 and 88.9 for the MD and HD datasets, respectively (see Appendix S2 for de novo results).

3.2 | Analysis of SNP datasets

We analysed the MD dataset in a phylogenetic context using Neighbour-Net, which nested all hybrids within a clade that also included pure *A. pulchellus* (Figure 4a). This nuclear SNP tree contrasts with mtDNA-based results, which grouped hybrids with *A. krugi* (Figure 4b; Jezkova et al., 2013a).

The PCA analysis of the MD and HD datasets yielded a signal of two population clusters along the first axis (corresponding to *A. pulchellus* and *A. krugi*; Figure 5a, Figure S3); these clusters explained 40.1% of the variance in the MD dataset and 49.7% of the variance in the HD dataset. *Anolis pulchellus* with native mtDNA (pure individuals) and with foreign mtDNA (hybrids) are differentiated along the second PCA axis, which explained 13.5% and 13.7% of the variance in the two datasets, respectively (see Appendix S2 for de novo results).

HI estimates for each dataset revealed that hybrid individuals displayed HIs that were distinct from (i.e., outside the range of) pure *A. pulchellus* (Figure 5b,c, Figure S4a,b), but that showed partial *A. krugi* ancestry (Table 1, Table S3; Figure 5b,c, Figure S4a,b). For

the MD dataset, these hybrids had average HI values ranging from 0.09 to 0.16. HI values based on the HD dataset for such hybrids were slightly higher, ranging from 0.21 to 0.22 (Table 1, Table S3; see Appendix S2 for de novo results). HI estimates at the chromosome level generally followed a similar pattern, with hybrids falling outside of the parental limits in the HD dataset (Figures S5 and S6). However, the HI estimates at the chromosome level in the MD dataset do not fall outside the parental limits because HI values derived from individual chromosomes displayed higher variance (indicated by larger confidence intervals; Figure S4). This finding is likely driven by the reduced number of chromosome-specific SNPs in the MD dataset (Figures S4 and S6).

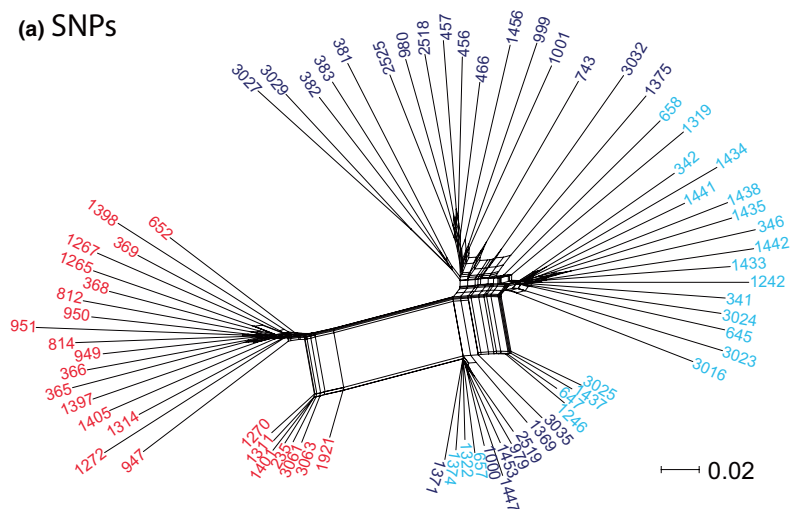
Differential introgression analysis of the MD dataset identified 367 and 591 candidate SNPs with a greater proportion of *A. krugi* and *A. pulchellus* alleles in hybrids than expected by chance, respectively (Figure 6; see Appendix S2 for de novo results). Demographic modelling for the MD dataset using *moments* identified the best model of diversification as the model with asymmetric gene flow between *A. pulchellus* and hybrid individuals and asymmetric gene flow between *A. krugi* and hybrids (Table S4). Migration rates into hybrids were estimated to be greater than migration from hybrids to either *A. pulchellus* or *A. krugi* (Table S4). Effective population size estimates were similar for *A. pulchellus* and hybrid populations and were smaller than the estimate for *A. krugi* (Table S4). The divergence between *A. krugi* and *A. pulchellus* was estimated to have occurred 1.29 million years ago (MYA), and the diversification into the modern populations was estimated to have occurred 0.98 MYA (Appendix S2).

Demographic simulations in *msprime* using *moments* parameter estimates produced 111,000 SNPs in two separate runs. The two runs averaged 9523 SNPs after filtering by *gghybrid*. The two simulated distributions were not significantly different ($D=0.012$, $p\text{-value}=0.54$). Simulation data exhibited a relatively uniform distribution of parental allele frequencies in hybrids, with some SNPs drifting to fixation, in comparison to our empirical data, which displayed a bimodal distribution. Kolmogorov-Smirnov tests indicated that the simulated and empirical distributions were significantly different for both runs (Run 1: $D=0.281$, $p\text{-value}<2.2\text{e-}16$; Run 2: $D=0.280$, $p\text{-value}<2.2\text{e-}16$; see Appendix S2 for de novo results; Figure S7).

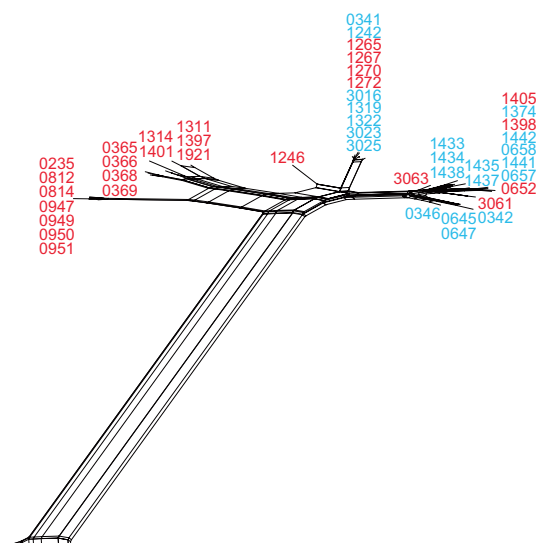
Differentially introgressed candidate SNPs with a greater proportion of *A. krugi* alleles in hybrids than expected by chance can be found throughout the *A. carolinensis* genome: on autosomes and sex chromosomes, as well as on unplaced scaffolds (Figure S8). We identified 97 reference SNPs that fell within the boundaries of annotated genes with known function, whereas 8 SNPs mapped to annotated genes with no known function. Thirty-six of these outlier SNPs associated with known genes are located in protein-coding regions of genes (i.e., exons); the other 69 are found in non-coding regions of the genes (i.e., introns; Table S5). The remaining outlier SNPs ($N=262$) map to non-coding regions. Candidate genes (linked to outlier SNPs) have functional annotations related to cellular metabolism, development, immunity and structural components within the

FIGURE 4 (a) Neighbour-Net trees depicting the genetic relationships among *Anolis pulchellus* (*A. pulchellus* with native mtDNA, dark blue), *A. krugi* (red) and hybrids (*A. pulchellus* with *A. krugi* mtDNA, light blue). The trees were inferred from the de novo clustering of (a) the medium-density SNP dataset including all individuals ($N=73$) and (b) the mtDNA dataset ($N=73$; Jezkova et al., 2013a). Sample numbers correspond to those in Table S1.

(a) SNPs



(b) mtDNA



cell (Table 2, Table S5). Three of the candidate genes that displayed a greater proportion of *A. krugi* alleles in hybrids, notch receptor 1 (*notch1*), delta-like canonical Notch ligand 1 (*dll1*) and jagged1 (*jag1*), are components of the Notch signalling pathway. This pathway is known to directly interact with the mitochondrial proteome by down-regulating key proteins in respiratory complex I and the glutamine catabolic process, which decreases dependence on exogenous glutamine for cell survival (Basak et al., 2014). Another candidate gene, NADH: Ubiquinone Oxidoreductase Subunit B11 (*ndufb11*), codes for a protein that is an accessory subunit of respiratory complex I (Stroud et al., 2016).

Differentially introgressed candidate SNPs with a greater proportion of *A. pulchellus* alleles in hybrids than expected by chance are also distributed throughout the *A. carolinensis* genome. We identified 84 reference SNPs that are within functionally annotated genes, whereas 7 SNPs map to annotated genes with unknown function. Twenty-six of the SNPs associated with known genes are located in protein-coding regions of candidate genes (i.e., exons); the other 59 are found in non-coding regions of genes (i.e., introns; Table S5). The remaining outlier SNPs map to non-coding regions ($N=506$). Candidate genes harbouring differentially introgressed

A. pulchellus alleles are related to cellular metabolism, development and structural components within the cell (Table S5). Two genes, acylglycerol kinase (*agk*) and *pwwp2b*, are known to be associated with thermoregulation (Prentki & Madiraju, 2008) and thermogenesis (Yan et al., 2021), respectively.

4 | DISCUSSION

Introgression can be an important source of adaptive genetic variation in nature (Hedrick, 2013). Adaptive introgression can introduce alleles associated with traits that increase individual fitness because they are better suited to the biotic and abiotic factors of a particular environment or associated with “repair” genes that have accumulated deleterious variants due to mutation and genetic drift. In this study, we developed an approach to identify patterns of differential introgression in naturally-occurring hybrids resulting from mosaic sympatry. Our approach leverages recent bioinformatic methods to identify parental source alleles (Bailey, 2020) and then employs a permutation procedure to determine whether the observed proportion of an allele from a particular parental population is greater in

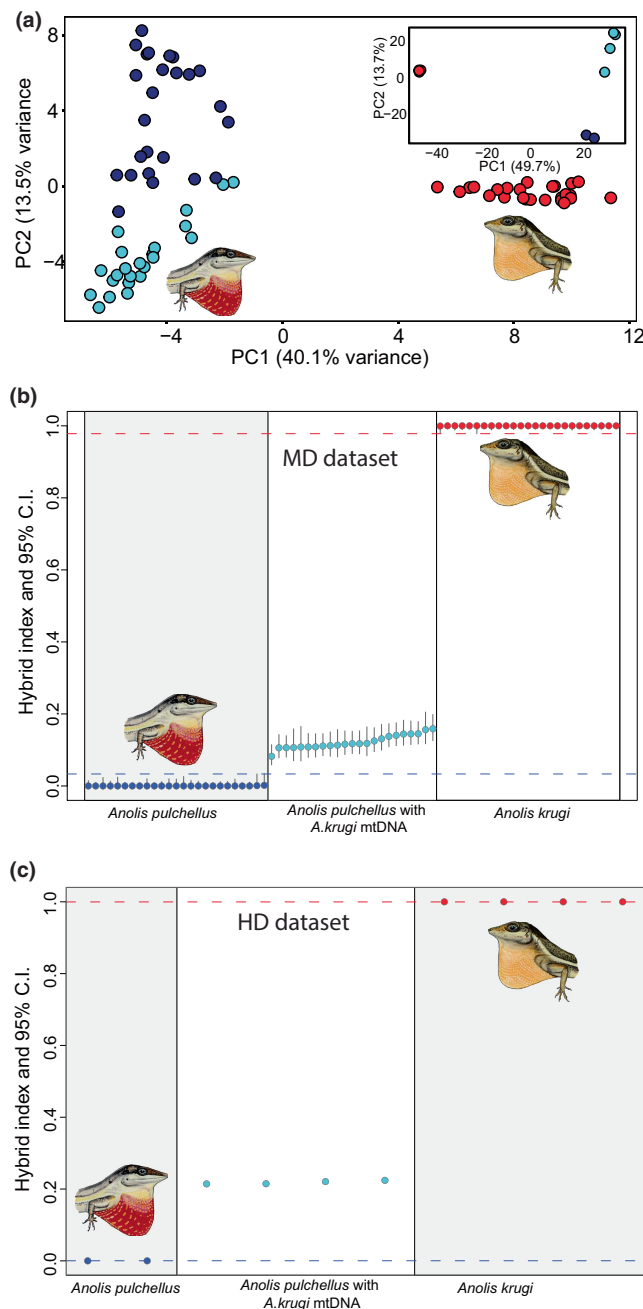


FIGURE 5 (a) Scatterplot of the first two axes of the principal component analysis (PCA) for *Anolis pulchellus* (*A. pulchellus* with native mtDNA, dark blue circles), *A. krugi* (red circles) and hybrids (*A. pulchellus* with *A. krugi* mtDNA, light blue circles). PC1 and PC2 were derived from the de novo clustering of the medium-density SNP dataset (N=73). Inset: Results of a PCA analysis of de novo clustering of the high-density SNP dataset (N=10). The percentage of variance explained by each axis is indicated in parentheses. (b, c) Hybrid index graphs with *A. pulchellus* (blue circles) and *A. krugi* (red circles) as parental references for the (b) medium-density and (c) high-density SNP datasets for hybrid *A. pulchellus* (*A. pulchellus* with *A. krugi* mtDNA, grey circles). Lines extending from each point are the 95% confidence interval for each individual. Dotted blue and red lines indicate the parental limit for each reference population.

hybrids than expected due to neutral introgression alone. This approach may be particularly useful in systems where the geography of introgression does not resemble a traditional clinal hybrid zone or where multiple independently localized instances of hybridization are detected.

Using our new locus permutation approach and coalescent simulations, we searched for SNPs exhibiting a signal of differential introgression, presumably due to selection, on the allelic content of hybrid genomes and linked these SNPs to known genes and their corresponding molecular functions. These candidate genes can thus be studied in detail in future experimental or comparative studies. Our approach enabled us to explore how differential introgression may have shaped the genomic ancestry and architecture of hybrid *A. pulchellus*, individuals that phenotypically resemble pure *A. pulchellus* but possess the mtDNA of *A. krugi*. This instance of natural hybridization and mitochondrial introgression is particularly interesting because i) documented cases of hybridization between sympatric anoles are remarkably rare in nature (Losos, 2009), ii) mitochondrial introgression is unidirectional, suggesting hybridization is dominated by matings between *A. pulchellus* males and *A. krugi* females and iii) mitochondrial introgression is geographically widespread, resulting in what appears to be a complete replacement of the native *A. pulchellus* mtDNA by that of *A. krugi* throughout a large area in western Puerto Rico (Jezkova et al., 2013a).

4.1 | Genomic composition of *A. pulchellus* hybrids

Genome-wide estimation of the ancestry of genetic markers in hybrid individuals revealed that hybrid genomes are predominately derived from *A. pulchellus* and that only 10%–20% of hybrid genomes come from *A. krugi* ancestry (Table 1; Figure 5b,c). This higher degree of *A. pulchellus* ancestry is consistent with the *pulchellus*-like phenotype of the hybrids and is qualitatively similar to patterns of uneven hybrid ancestry observed in other taxa (Llopart et al., 2005; Rieseberg et al., 1996). Typically, reduced ancestry from one parental lineage can be attributed to one of three, non-mutually exclusive processes: (i) continuous backcrossing of hybrids with one of the parental lineages (Bachtrog et al., 2006; Rand et al., 2006), (ii) restricted introgression that results in underrepresentation of one of the parental genomes in the hybrids due to intrinsic factors, such as chromosomal incompatibilities or strong epistatic interactions in the nuclear genome (Llopart et al., 2005, 2014; Rieseberg, 2011; Rieseberg et al., 1996) and (iii) subsequent selection acting in favour or against one or more parental alleles in the hybrids, and backcrosses due to extrinsic factors, such as environmental conditions. While our data do not allow us to explicitly test among these three processes, below we discuss expectations for each process and discuss whether they are consistent with our results.

Backcrossing is expected to reduce the fraction of *A. krugi* ancestry in the genome of hybrids over generations (cf. Bachtrog et al., 2006; Rand et al., 2006). For neutral loci (i.e., freely introgressing alleles),

TABLE 1 Average hybrid index (HINDEX) estimates, 95% confidence interval (in parentheses) for *Anolis pulchellus*, hybrids (*A. pulchellus* with *A. krugi* mtDNA), and *A. krugi*, the parental limit for each source population (*A. pulchellus* and *A. krugi*) for the medium- and high-density SNP reference datasets, and the number of SNPs included in each analysis.

Dataset	<i>Anolis pulchellus</i>	Hybrids	<i>Anolis krugi</i>	Parental limits (<i>A. pulchellus</i> , <i>A. krugi</i>)	SNPs
Medium-density SNP reference	0.001 (0–0.016)	0.122 (0.092–0.161)	0.999 (0.997–1)	0.0145, 0.997	2188
High-density SNP reference	0 (0–0.002)	0.219 (0.211–0.226)	0.999 (0.998–1)	0.003, 0.998	6606

Note: HINDEX represents an estimate of the proportion of alleles that were inherited from one of the two parental species, and in this study, HINDEX is an estimate of the proportion of alleles that were inherited from *A. krugi*. Parental limits are used to determine whether an individual exhibits signals of hybridization. An individual that falls outside of the parental limits can be considered a hybrid.

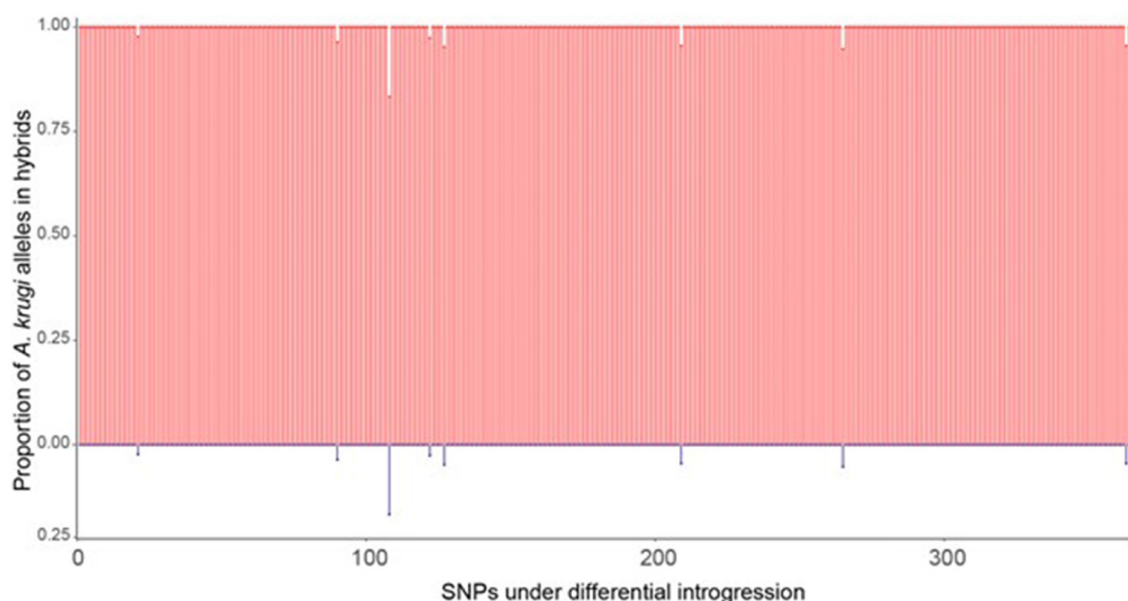


FIGURE 6 The proportion of *Anolis krugi* (red) and *A. pulchellus* (blue) alleles in hybrids at the 367 candidate SNPs exhibiting a signal of differential introgression in the medium-density SNP reference dataset.

backcrossing should also lead to variation across hybrids in both the number and genomic locations of *A. krugi* alleles. This is because, in the absence of selection, backcrossing should result in stochastic inheritance of parental alleles and because different individuals may be the product of different numbers of backcrossing events with unique outcomes and distinct recombination events. Our results suggest that backcrossing alone does not adequately explain the reduced representation of *A. krugi* alleles in the hybrids. First, all hybrids exhibited a similar proportion (10%–20%) of introgressed *A. krugi* loci. Second, all our analyses suggest the presence of differentially introgressed alleles in the genomes of the hybrids. In particular, the coalescent simulations suggested that the empirical frequencies of differentially introgressed alleles differed significantly from the simulated frequencies, as many *A. pulchellus* and *A. krugi* alleles are present at higher frequencies in hybrids than what would be expected at random. Indeed, only ca. 5% of *A. krugi* loci exhibited a signal of random introgression in the hybrid genomes. Although this percentage was higher in the de novo assembly (57% of de novo SNPs showed a signal of random introgression; Appendix S2), our findings indicate that a large proportion of the *A. krugi* alleles in the hybrids exhibit differential introgression. We believe that the discordance between inferences from the reference

genome and de novo inferences may be due to the loci that exhibit random introgression representing neutral loci, which generally accumulate substitutions faster than non-neutral loci and would therefore be less likely to map to the reference genome of a different species. The higher number of randomly introgressed loci in the de novo dataset can also be caused by uncorrected LD, given that the location of loci in the genome is unknown.

Restricted introgression may also lead to a reduced representation of the genome of one of the parental species in the hybrids. Restricted introgression can occur due to intrinsic factors that limit recombination, such as the presence of chromosomal divergence, non-linear linkage blocks (which prevent recombination), deleterious interspecific interactions between loci (Aeschbacher et al., 2017; Llopart et al., 2005, 2014; Martin et al., 2019; Rieseberg, 2011; Schumer et al., 2018) or broad selection against minor-parent ancestry (in a genome background dominated by another parental species) due to disruptions in epistatic interactions among coevolved loci from the major parental lineage. Restricted introgression is typically observed in deeply divergent species pairs, as greater divergence times increase the probability of chromosomal incompatibilities arising between hybridizing species (Ayala et al., 1974; Harrison

TABLE 2 Gene ontology (GO) information for the 11 candidate, differentially introgressed SNPs discussed in Sections 3.2 and 4.2.

Dataset	SNP location	Gene name	Genomic element	GO categories
Reference	chr1_134137907	Jagged1 (<i>jag1</i>)	Intron	F: Notch binding, C: adherens junction, C: apical plasma membrane, P: positive regulation of Notch signalling pathway, P: glomerular visceral epithelial cell development
Reference	chr1_223343339	Delta-like canonical notch ligand 1 (<i>dll1</i>)	Exon	F: calcium ion binding; F: notch binding; P: astrocyte development; P: cell differentiation
Reference	chr11_40594200	Spectrin beta, non-erythrocytic 5 (<i>sptbn5</i>)	Intron	C: cytoskeleton, C: cytoplasm, P: actin filament capping, F: actin binding, F: myosin tail binding
Reference	chr2_68882726	NADH:ubiquinone oxidoreductase subunit B11 (<i>ndufb11</i>)	Intron	P: mitochondrial respiratory chain complex I assembly, C: mitochondrial respiratory chain complex I, C: mitochondrial respiratory chain complex I, C: membrane, C: integral component of membrane
Reference	chr3_170687954	Cbl proto-oncogene B (<i>cblb</i>)	Intron	P: immune response, C: membrane raft, P: negative regulation of epidermal growth factor-activated receptor activity, P: signal transduction, F: receptor tyrosine kinase binding
Reference	chr6_5937789	Sonic hedgehog signalling molecule (<i>shh</i>)	Exon	F: calcium ion binding; P: androgen metabolic process, P: animal organ formation
Reference	AAWZ02036232_16058	Notch receptor 1 (<i>notch1</i>)	Exon	F: calcium ion binding; F: chromatin DNA binding; P: animal organ regeneration; P: aortic valve morphogenesis
Reference	GL343225.1_893596	BARX homeobox 1 (<i>barx1</i>)	Exon	F: DNA-binding transcription factor; P: anterior/posterior pattern specification, P: digestive system development
Reference	GL343573.1_277896	Aldehyde Dehydrogenase 1 Family Member A2 (<i>aldh1a2</i>)	Intron	F: 3-chloroallyl aldehyde dehydrogenase activity; P: blood vessel development; P: cardiac muscle tissue development
de novo	dDocent_Contig_2381	Asunder spermatogenesis regulator (<i>asun</i>)	4E-39	P: cell division, P: centrosome localization
de novo	dDocent_Contig_3278	Gamma-Aminobutyric Acid Type A Receptor Subunit Alpha5 (<i>gabra5</i>)	3E-35	F: GABA-A receptor activity; P: associative learning; P: brain development; P: nervous system process

Note: Candidates were found to be located within a gene transcript in the genome of *Anolis carolinensis* (see Section 2.3 for detailed methods; Alföldi et al., 2011). For the nine reference candidates, columns represent the SNP location on the *A. carolinensis* assembly, the gene name, the genomic element where the SNP is located (i.e., intron, exon) and the GO categories for each gene, which can provide information about their biological relevance (i.e., cellular component (C), molecular function (F), biological process (P)). Reference SNPs are named based on the chromosome or unplaced scaffold where they are located (characters before the underscore) and their position on that chromosome or unplaced scaffold (characters after the underscore). For the two de novo candidates, columns represent the dDocent contig that candidates are located on, the gene name, the expected value (E-value) from BLAST (Altschul et al., 1990) and the GO categories for each gene (see Appendix S1 for methods used to annotate de novo candidate SNPs). The GO information for all candidate SNPs identified as exhibiting differential introgression appears in Table S5.

& Larson, 2014). In closely related species, such as the sister taxa *A. krugi* and *A. pulchellus* (these anoles have the same karyotype, $N=29$, with 13 pairs of autosomes and 3 unpaired sex chromosomes; Gorman & Atkins, 1969; Gorman & Stamm, 1975), such chromosomal incompatibilities should be less common, although they may still exist. Experimental studies in other systems suggest that when restricted introgression occurs, hybrids exhibit 5%–10% of genomic markers from one of the parental species (Llopart et al., 2005, 2014; Rieseberg, 2011; Rieseberg et al., 1996). Hybridization models also predict that a large proportion of the same introgressed loci are found across independent hybridization events (Rieseberg, 2011; Rieseberg et al., 1996). These loci will therefore exhibit a signal of differential introgression. Indeed, the similar proportion of *A. krugi* ancestry in the nuclear genomes of the hybrids, together with the large proportion of loci exhibiting differential introgression (43%–95%), are consistent with the restricted introgression model. However, more definitive evidence for restricted introgression would be provided by finding differentially introgressed loci in the F2 generation of hybrids.

Finally, the occurrence of differentially introgressed loci together with a complete or near-complete replacement of pure *A. pulchellus* by hybrids across a large geographic area of Puerto Rico suggests a scenario in which natural selection favours certain combinations of *A. pulchellus* and *A. krugi* loci in particular environments. The selective pressures underlying differential introgression can be diverse, ranging from purifying selection that removes accumulated deleterious alleles to positive selection, where introgressed alleles provide a fitness advantage to hybrids (Ballard & Whitlock, 2004; Harrison & Larson, 2016; Hill et al., 2019). We elaborate on these points below.

4.2 | Loci with evidence for differential introgression

A primary motivation for this study was to assess whether at least some *A. krugi* loci in the genome of hybrids show a signal of differential introgression. These loci should occur in the hybrids at a higher frequency than expected by chance. Although some SNPs did drift to fixation in our coalescent simulations, the number of SNPs found at high frequencies in the hybrids was significantly higher in the empirical data than in our simulations. Although we cannot rule out that model under-fitting due to the simplicity of our demographic models could bias our results (e.g., no bottlenecks of hybrid populations were considered), the differences between empirical and simulated data suggest that drift alone is unlikely to explain the observed pattern of introgression in hybrids and that selection has likely played a role in establishing the genomic composition of *A. pulchellus* hybrids.

We found 367 SNPs that exhibit differential introgression in our permutation approach, implying that hybrids originating from independent hybridization events possess a large proportion of the same genomic regions as *A. krugi*. In general, candidate SNPs identified by the permutation approach displayed a high frequency of *A. krugi* alleles in the hybrids. However, whether an SNP is identified as a

candidate is influenced by allele sharing between parental populations (i.e., parental polymorphism). In particular, the permutation procedure identifies SNPs as candidates when the *A. krugi* allele is close to fixation in hybrid individuals but at low frequency in *A. pulchellus* individuals. Conversely, SNPs identified as non-candidates exhibit a higher frequency of *A. krugi* alleles in both parental populations. This results in permuted hybrids harbouring a higher proportion of *A. krugi* alleles than empirical hybrids, leading to elevated p-values. The Bonferroni method to correct for multiple tests further decreases the false positive rate but can also inflate the false negative rate (Type II error; Verhoeven et al., 2005) by removing some SNPs from the candidate list. Importantly, each candidate SNP identified here should be investigated further, as our permutation method does not explicitly test for the effect of genetic drift.

Of the 367 SNPs exhibiting a signal of differential introgression, 97 are located within known gene transcripts that are involved in critical cellular, physiological and developmental processes (e.g., *dll1*; cellular development; *notch1*; cell signalling), or are structural cellular components (e.g., Spectrin beta chain, non-erythrocytic 5: *sptbn5*; cytoskeleton component; Table 2, Table S5). In our study, the proportion of candidate SNPs that are located within a gene transcript (28.6%) falls within the range of figures reported by other studies that sought to identify genomic regions under selection. Those reports identified between 8% and 73.6% candidate loci that mapped to known DNA/protein sequence identifiers (Byer et al., 2021; Dalongeville et al., 2018; Tigano et al., 2017).

Previous research suggests that both purifying and positive selection can drive the observed patterns of differentially introgressed loci. Purifying selection removes deleterious alleles to preserve the proper functions of genes and the interactions of those genes in their respective pathways (Cvijović et al., 2018). Purifying selection has been primarily believed to act against introgression (López-Delgado et al., 2020; Petr et al., 2019), but can also act in favour of introgression of foreign mtDNA and associated mito-nuclear interactions (Hill et al., 2019; Llopart et al., 2014; Sloan et al., 2017), such as the Notch signalling pathway; see Section 4.3 below).

Differential introgression can also be driven by positive selection, where introgressed alleles provide an advantage to hybrids (Hedrick, 2013). Differential introgression has been linked to adaptation to local environments, including environmental temperatures (Harrison & Larson, 2016). A recent study of North American box turtles (*Terrapene* spp.; Martin et al., 2020) identified signatures of differential introgression associated with temperature and implicated genes involved in temperature-dependent sex determination, immune function and molecular pathways responsible for the development of skeletal, muscle and nervous tissues that play a role in tolerance to anoxia and hypoxia. Many of the candidate genes associated with differentially introgressed loci that we identified in *A. pulchellus* hybrids serve functional roles similar to the roles of the candidate genes identified in box turtles (Martin et al., 2020): BARX Homeobox1 (*barx1*), aldehyde dehydrogenase 1 family member A2 (*aldh1a2*), asunder spermatogenesis regulator (*asun*) and sonic hedgehog (*shh*) are involved in development; Cbl proto-oncogene B

(*cblb*) is involved in immune function; Gamma-aminobutyric acid receptor subunit alpha-5 (*gabrb5*) is involved in the development of the nervous tissue; and acylglycerol kinase (*agk*) is involved in thermoregulation (Table 2, Table S5).

The contrasting thermal preferences of *A. pulchellus* and *A. krugi* are likely to impose distinct thermal constraints on these anoles. As previously stated, *A. krugi* prefers partially shaded, cooler habitats, whereas *A. pulchellus* prefers open, sunnier, warmer habitats. In anoles and other lizards, species that inhabit partially shaded conditions tend to be active at cooler temperatures, which likely influences their performance curves (Gunderson & Leal, 2016). *Anolis pulchellus* hybrids may therefore exhibit a performance curve with a "broader shoulder" and reach optimal performance at lower temperatures. If so, this thermal profile may allow hybrids to outperform pure *A. pulchellus* at cooler temperatures. A broad shoulder performance curve can also increase the daily activity times of hybrids by allowing them to be active in early morning and/or late afternoon periods during which ambient temperatures tend to be cooler. For ectotherms, total activity time is thought to be critical for population persistence (Gunderson & Leal, 2016; Huey et al., 2010).

Candidate genes associated with differentially introgressed loci in *A. pulchellus* have been previously implicated in limb development, in particular the sonic hedgehog gene (*shh*; Tickle & Towers, 2017) and genes associated with Notch signalling (Mašek & Andersson, 2017). Limb length is an adaptive trait in anoles (Donihue et al., 2018; Losos et al., 1997), and a recent study of *A. sagrei* identified signals of positive selection in a genomic region that contains several candidate genes that are involved in limb development (Bock et al., 2021). Both the forelimbs and hind limbs of *A. krugi* are longer than those of *A. pulchellus* (Losos, 1990), and in anoles, longer limbs provide enhanced maximal sprint speed, which permits rapid locomotion to capture prey and escape predators (Losos & Irschick, 1996), a trait that may be selectively advantageous. We currently lack morphological data on *A. pulchellus* hybrids, and it will be informative to determine whether limb length differs between pure *A. pulchellus* and hybrid individuals. Similarly, additional work is needed to understand how the Notch pathway impacts limb development in anoles. Together, our findings suggest that the observed signatures of differential introgression in *A. pulchellus* hybrids may be caused by selection on physiological and morphological traits. Still, we recognize that genes we identified to exhibit differential introgression may not be under selection themselves but instead be linked to genes under selection that were not represented in our functional inferences.

4.3 | Mitochondrial-nuclear interactions

Mitochondrial-nuclear (mito-nuclear) interactions are fundamental to most eukaryotes, and many biological processes, such as energy production, cellular respiration, cellular development and regulation, hinge on these coordinated actions between genes of the nuclear and mitochondrial genomes (Currat et al., 2008; Hedrick, 2013; Toews & Brelsford, 2012; Wolff et al., 2014). Mito-nuclear genomes

of non-avian reptiles encode 13 protein-coding genes, but the function of mitochondria can depend on >1000 different proteins encoded by mitochondrially targeted genes in the nucleus (Sloan et al., 2017). *Anolis krugi*'s mitochondrial introgression into *A. pulchellus* was previously attributed to selection on the mitochondrial genome (Jezkova et al., 2013a). Adaptive introgression of foreign mtDNA may occur in species or populations that have suffered from the accumulation of deleterious mitochondrial mutations (Llopart et al., 2014; Sloan et al., 2017), although adaptive introgression has also been linked to thermal tolerance and local adaptation to different climatic conditions (Sloan et al., 2017; Toews & Brelsford, 2012).

Recent studies have shown that mitochondrial introgression may be accompanied by preferential introgression of regions that are enriched for nuclear-encoded mitochondrial targeted genes (Beck et al., 2015; Morales et al., 2017), and mito-nuclear co-introgression has been indirectly inferred to affect basal metabolic rate (Boratyński et al., 2016). Our results contribute to this growing set of examples and indicate that differential introgression at some nuclear loci in *A. pulchellus* hybrids may be driven by selection to maintain functional mitochondrial-nuclear interactions with *A. krugi*'s mtDNA genome. The Notch signalling pathway, which is associated with some of the differentially introgressed SNPs in hybrid *A. pulchellus*, is an evolutionary conserved pathway (Shi & Wang, 2017) that is known to affect the mitochondrial proteome and is crucial for cellular development (Bray, 2006) and metabolism (Basak et al., 2014). The three genes involved in Notch signalling that are differentially introgressed in *A. pulchellus* hybrids encode proteins that represent the major components of this pathway (receptor; *notch1* and ligands; *dll1*, *jag1*; Shi & Wang, 2017). This finding suggests that Notch signalling may be important in our study system and that differential introgression may be driven by the need to preserve or optimize this pathway (Hill et al., 2019; Wolff et al., 2014). Specifically, the activation of *notch1* alters 10 unique proteins by down-regulating protein expression in respiratory complex I and the glutamine catabolic process; 8 of those proteins belong to mitochondria-localized metabolic pathways, including oxidative phosphorylation, glutamine metabolism, the Krebs cycle and fatty acid oxidation (Basak et al., 2014).

Metabolism is critical for cellular function and generally increases with temperature, as higher temperatures require more adenosine triphosphate production, thus increasing metabolism (Clarke & Fraser, 2004). This may be particularly important in our system, as ectothermic vertebrates, such as *Anolis* lizards, display a strong reliance on environmental thermoregulation (Martin et al., 2020). The Notch signalling pathway also affects the regeneration and metabolism of nicotinamide adenine dinucleotide + hydrogen (NADH) and nicotinamide adenine dinucleotide phosphate (NADPH), and respiratory complex I in general, the first and largest component of the mitochondrial oxidative phosphorylation system (Brandt, 2006). This too may be important in our system, as respiratory complex I plays a central role in energy metabolism and overall respiration (Sharma et al., 2009). Therefore, alterations to the function or composition of respiratory complex I may affect metabolism and cellular respiration, which in turn may influence the fitness of *A. pulchellus* hybrids. Our

preliminary findings suggest the potential for patterns of differential introgression to be linked to selection in mito-nuclear interactions, but this intriguing scenario requires confirmation, perhaps by performing selective experiments in the wild.

5 | CONCLUSION

We identified evidence of neutral and differential introgression in natural hybrids of the Caribbean anoles *Anolis pulchellus* and *A. krugi* in Puerto Rico. Our sampling of nuclear SNPs supports the previous inference of hybridization between these two sister species based on mitochondrial data alone (Jezkova et al., 2013a). The nuclear genomic composition of the hybrids is dominated by loci of *A. pulchellus* ancestry, and the phenotype of these hybrids is indistinguishable from parental *A. pulchellus*. Still, a small but consistent proportion (10%–20%) of the hybrid genomes are of *A. krugi* ancestry. In these hybrids, a large proportion (43%–95%) of introgressed *A. krugi* alleles display signatures of differential introgression, and these alleles are associated with genes involved in key mitochondrial and developmental pathways, suggesting selection acting on the hybrids that preserves or optimizes mitochondrial-nuclear interactions between *A. krugi* mitochondrial genomes and *A. krugi* nuclear-encoded alleles. Our findings highlight the important role of variation in genetic introgression in shaping the nuclear genomic composition of hybrids. Further investigation of alleles exhibiting signatures of differential introgression and the functional relevance and ramifications of differentially introgressed loci may lead to a better understanding of hybridization and its underlying mechanisms in this system and of the possible effect of mitochondrial-nuclear interactions on the phenotype.

AUTHOR CONTRIBUTIONS

K.F., T.A.C., T.J. and J.A.R.-R. designed the study. J.A.R.-R. conducted the field work. K.F., A.A., M.F., D.R.S., D.C.C. and T.J. performed the analyses. K.F. and T.J. wrote the first draft of the manuscript; K.F., T.A.C., M.L., T.J. and J.A.R.-R. provided the funding. All authors contributed to writing and editing the manuscript.

ACKNOWLEDGEMENTS

We thank Jill Castoe for assistance in the laboratory; Andor Kiss of the Center for Bioinformatics & Functional Genomics (CBFG) at Miami University for instrumentation and computational support; Jens Mueller of the Research Computing group at Miami University for computational support and invaluable advice with the Miami RedHawk Cluster; Jon Puritz for helpful advice with dDocent; Richard Bailey for guidance in utilizing gghybrid. We also thank Simon Martin for insightful suggestions and guidance with msprime. This project was supported by Miami University start-up funds to T.J.; by a National Science Foundation Graduate Research Fellowship Program to K.F. (Award #2037786); by an NSF grant to M.L. (IOS-1051793); by faculty start-up funds from the University of Texas, Arlington and NSF support (DEB-1655571) to T.A.C.; and

by institutional funds for research support from the University of Nevada, Las Vegas to J.A.R.-R.

CONFLICT OF INTEREST STATEMENT

The authors report no conflict of interest.

DATA AVAILABILITY STATEMENT

Data and analytical scripts are available on Dryad at <https://doi.org/10.5061/dryad.cz8w9gj8v>. HybridFindR is available at <https://github.com/kfarleigh/HybridFindR>. Processed and demultiplexed RADseq data are available at NCBI BioProject PRJNA1023949.

ORCID

Keaka Farleigh  <https://orcid.org/0000-0002-9195-121X>
 Alfredo Ascanio  <https://orcid.org/0000-0001-9987-7977>
 Mackenzie E. Farleigh  <https://orcid.org/0000-0003-3402-9728>
 Drew R. Schield  <https://orcid.org/0000-0001-7854-9480>
 Daren C. Card  <https://orcid.org/0000-0002-1629-5726>
 Todd A. Castoe  <https://orcid.org/0000-0002-5912-1574>
 Tereza Jezkova  <https://orcid.org/0000-0003-4114-4564>
 Javier A. Rodríguez-Robles  <https://orcid.org/0000-0003-4501-9875>

REFERENCES

- Abbott, R., Albach, D., Ansell, S., Arntzen, J. W., Baird, S. J. E., Bierne, N., Boughman, J., Brelsford, A., Buerkle, C. A., Buggs, R., Butlin, R. K., Dieckmann, U., Eroukhanoff, F., Grill, A., Cahan, S. H., Hermansen, J. S., Hewitt, G., Hudson, A. G., Jiggins, C., ... Zinner, D. (2013). Hybridization and speciation. *Journal of Evolutionary Biology*, 26(2), 229–246. <https://doi.org/10.1111/j.1420-9101.2012.02599.x>
- Aeschbacher, S., Selby, J. P., Willis, J. H., & Coop, G. (2017). Population-genomic inference of the strength and timing of selection against gene flow. *Proceedings of the National Academy of Sciences of the United States of America*, 114(27), 7061–7066. <https://doi.org/10.1073/pnas.1616755114>
- Álföldi, J., Di Palma, F., Grabherr, M., Williams, C., Kong, L., Mauceli, E., Russell, P., Lowe, C. B., Glor, R. E., Jaffe, J. D., Ray, D. A., Boissinot, S., Shedlock, A. M., Botka, C., Castoe, T. A., Colbourne, J. K., Fujita, M. K., Moreno, R. G., Ten Hatters, B. F., ... Lindblad-Toh, K. (2011). The genome of the green anole lizard and a comparative analysis with birds and mammals. *Nature*, 477(7366), 587–591. <https://doi.org/10.1038/nature10390>
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215(3), 403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
- Anderson, E. (1949). *Introgressive hybridization*. John Wiley and Sons.
- Anderson, E., & Hubricht, L. (1938). Hybridization in Tradescantia. III. The evidence for introgressive hybridization. *American Journal of Botany*, 25(6), 396–402. <https://doi.org/10.1002/j.1537-2197.1938.tb09237.x>
- Ayala, F. J., Tracey, M. L., Hedgecock, D., & Richmond, R. C. (1974). Genetic differentiation during the speciation process in *Drosophila*. *Evolution*, 28(4), 576–592. <https://doi.org/10.2307/2407283>
- Bachtrog, D., Thornton, K., Clark, A., & Andolfatto, P. (2006). Extensive introgression of mitochondrial DNA relative to nuclear genes in the *Drosophila yakuba* species group. *Evolution*, 60(2), 292–302. <https://doi.org/10.1554/05-337.1>
- Bailey, R. (2020). *gghybrid: R package for evolutionary analysis of hybrids and hybrid zones* (1.0.0). <https://doi.org/10.5281/zenodo.3676499>

- Baird, N. A., Etter, P. D., Atwood, T. S., Currey, M. C., Shiver, A. L., Zachary, A., Selker, E. U., Cresko, W. A., & Johnson, E. A. (2008). Rapid SNP discovery and genetic mapping using sequenced RAD markers. *PLoS One*, 3(10), e3376. <https://doi.org/10.1371/journal.pone.0003376>
- Ballard, J. W. O., & Melvin, R. G. (2010). Linking the mitochondrial genotype to the organismal phenotype: Invited review. *Molecular Ecology*, 19(8), 1523–1539. <https://doi.org/10.1111/j.1365-294X.2010.04594.x>
- Ballard, J. W. O., & Whitlock, M. C. (2004). The incomplete natural history of mitochondria. *Molecular Ecology*, 13(4), 729–744. <https://doi.org/10.1046/j.1365-294X.2003.02063.x>
- Barton, N. H. (2001). The role of hybridization in evolution. *Molecular Ecology*, 10(3), 551–568. <https://doi.org/10.1046/j.1365-294X.2001.01216.x>
- Basak, N. P., Roy, A., & Banerjee, S. (2014). Alteration of mitochondrial proteome due to activation of Notch1 signaling pathway. *Journal of Biological Chemistry*, 289(11), 7320–7334. <https://doi.org/10.1074/jbc.M113.519405>
- Baumdicker, F., Bisschop, G., Goldstein, D., Gower, G., Ragsdale, A. P., Tsambos, G., Zhu, S., Eldon, B., Ellerman, E. C., Galloway, J. G., Gladstein, A. L., Gorjanc, G., Guo, B., Jeffery, B., Kretzschmar, W. W., Lohse, K., Matschiner, M., Nelson, D., Pope, N. S., ... Kelleher, J. (2022). Efficient ancestry and mutation simulation with msprime 1.0. *Genetics*, 220(3), iyab229. <https://doi.org/10.1093/genetics/iyab229>
- Beck, E. A., Thompson, A. C., Sharbrough, J., Brud, E., & Llopart, A. (2015). Gene flow between *Drosophila yakuba* and *Drosophila santomea* in subunit V of cytochrome c oxidase: A potential case of cytonuclear cointrogression. *Evolution*, 69(8), 1973–1986. <https://doi.org/10.1111/evo.12718>
- Binns, D., Dimmer, E. C., Huntley, R. P., Barrell, D. G., O'Donovan, C., & Apweiler, R. (2009). QuickGO: A web-based tool for gene ontology searching. *Bioinformatics*, 25(22), 3045–3046. <https://doi.org/10.1093/bioinformatics/btp536>
- Bock, D. G., Baeckens, S., Pita-Aquino, J. N., Chejanovski, Z. A., Michaelides, S. N., Muralidhar, P., Lapiedra, O., Park, S., Menke, D. B., Geneva, A. J., Losos, J. B., & Kolbe, J. J. (2021). Changes in selection pressure can facilitate hybridization during biological invasion in a Cuban lizard. *Proceedings of the National Academy of Sciences of the United States of America*, 118(42), e2108638118. <https://doi.org/10.1073/pnas.2108638118>
- Boratyński, Z., Ketola, T., Koskela, E., & Mappes, T. (2016). The sex specific genetic variation of energetics in bank voles, consequences of introgression? *Evolutionary Biology*, 43(1), 37–47. <https://doi.org/10.1007/s11692-015-9347-2>
- Boratyński, Z., Melo-Ferreira, J., Alves, P. C., Berto, S., Koskela, E., Pentikäinen, O. T., Tarroso, P., Ylilauri, M., & Mappes, T. (2014). Molecular and ecological signs of mitochondrial adaptation: Consequences for introgression? *Heredity*, 113(4), 277–286. <https://doi.org/10.1038/hdy.2014.28>
- Bourgeois, Y., Ruggiero, R. P., Manthey, J. D., & Boissinot, S. (2019). Recent secondary contacts, linked selection, and variable recombination rates shape genomic diversity in the model species *Anolis carolinensis*. *Genome Biology and Evolution*, 11(7), 2009–2022. <https://doi.org/10.1093/gbe/evz110>
- Brandt, U. (2006). Energy converting NADH: Quinone oxidoreductase (Complex I). *Annual Review of Biochemistry*, 75(1), 69–92. <https://doi.org/10.1146/annurev.biochem.75.103004.142539>
- Bray, S. J. (2006). Notch signalling: A simple pathway becomes complex. *Nature Reviews Molecular Cell Biology*, 7(9), 678–689. <https://doi.org/10.1038/nrm2009>
- Bryant, D., & Moulton, V. (2004). Neighbor-net: An agglomerative method for the construction of phylogenetic networks. *Molecular Biology and Evolution*, 21(2), 255–265. <https://doi.org/10.1093/molbev/msh018>
- Buerkle, C. A. (2005). Maximum-likelihood estimation of a hybrid index based on molecular markers. *Molecular Ecology Notes*, 5(3), 684–687. <https://doi.org/10.1111/j.1471-8286.2005.01011.x>
- Burnham, K. P., & Anderson, D. R. (2004). Multimodel inference: Understanding AIC and BIC in model selection. *Sociological Methods & Research*, 33(2), 261–304. <https://doi.org/10.1177/0049124104268644>
- Burton, R. S. (2022). The role of mitonuclear incompatibilities in allopatric speciation. *Cellular and Molecular Life Sciences*, 79(2), 103. <https://doi.org/10.1007/s00018-021-04059-3>
- Burton, R. S., & Barreto, F. S. (2012). A disproportionate role for mtDNA in Dobzhansky–Muller incompatibilities? *Molecular Ecology*, 21(20), 4942–4957. <https://doi.org/10.1111/mec.12006>
- Byer, N. W., Holding, M. L., Crowell, M. M., Pierson, T. W., Dilts, T. E., Larrucea, E. S., Shoemaker, K. T., & Matocq, M. D. (2021). Adaptive divergence despite low effective population size in a peripherally isolated population of the pygmy rabbit, *Brachylagus idahoensis*. *Molecular Ecology*, 30(17), 4173–4178. <https://doi.org/10.1111/mec.16040>
- Capblancq, T., Mavárez, J., Rioux, D., & Després, L. (2019). Speciation with gene flow: Evidence from a complex of alpine butterflies (*Coenonympha*, Satyridae). *Ecology and Evolution*, 9(11), 6444–6457. <https://doi.org/10.1002/ece3.5220>
- Catchen, J., Hohenlohe, P. A., Bassham, S., & Amores, A. (2013). Stacks: An analysis tool set for population genomics. *Molecular Ecology*, 22(11), 3124–3140. <https://doi.org/10.1111/mec.12354>
- Chang, C. C., Rodriguez, J., & Ross, J. (2016). Mitochondrial–nuclear epistasis impacts fitness and mitochondrial physiology of interpopulation *Caenorhabditis briggsae* hybrids. *G3: Genes, Genomes, Genetics*, 6(1), 209–219. <https://doi.org/10.1534/g3.115.022970>
- Clarke, A., & Fraser, K. P. (2004). Why does metabolism scale with temperature? *Functional Ecology*, 18(2), 243–251. <https://doi.org/10.1111/j.0269-8463.2004.00841.x>
- Curat, M., Ruedi, M., Petit, R. J., & Excoffier, L. (2008). The hidden side of invasions: Massive introgression by local genes. *Evolution*, 62(8), 1908–1920. <https://doi.org/10.1111/j.1558-5646.2008.00413.x>
- Cvijović, I., Good, B. H., & Desai, M. M. (2018). The effect of strong purifying selection on genetic diversity. *Genetics*, 209(4), 1235–1278. <https://doi.org/10.1534/genetics.118.301058>
- Dalongeville, A., Benestan, L., Mouillot, D., Lobreaux, S., & Manel, S. (2018). Combining six genome scan methods to detect candidate genes to salinity in the Mediterranean striped red mullet (*Mullus surmuletus*). *BMC Genomics*, 19(217), 1–13. <https://doi.org/10.1186/s12864-018-4579-z>
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., Handsaker, R. E., Lunter, G., Marth, G. T., Sherry, S. T., McVean, G., & Durbin, R. (2011). The variant call format and VCFtools. *Bioinformatics*, 27(15), 2156–2158. <https://doi.org/10.1093/bioinformatics/btr330>
- Doiron, S., Bernatchez, L., & Blier, P. U. (2002). A comparative mitogenomic analysis of the potential adaptive value of arctic charr mtDNA introgression in brook charr populations (*Salvelinus fontinalis* Mitchell). *Molecular Biology and Evolution*, 19(11), 1902–1909. <https://doi.org/10.1093/oxfordjournals.molbev.a004014>
- Donihue, C. M., Herrel, A., Fabre, A.-C., Kamath, A., Geneva, A. J., Schoener, T. W., Kolbe, J. J., & Losos, J. B. (2018). Hurricane-induced selection on the morphology of an Island lizard. *Nature*, 560(7716), 88–91. <https://doi.org/10.1038/s41586-018-0352-3>
- Dunn, O. J. (1961). Multiple comparisons among means. *Journal of the American Statistical Association*, 56(293), 52–64. <https://doi.org/10.1080/01621459.1961.10482090>
- Emerson, K. J., Merz, C. R., Catchen, J. M., Hohenlohe, P. A., Cresko, W. A., Bradshaw, W. E., & Holzapfel, C. M. (2010). Resolving postglacial phylogeography using high-throughput sequencing. *Proceedings of the National Academy of Sciences of the United States of America*, 107(1), 104–109. <https://doi.org/10.1073/pnas.0906040107>

- States of America*, 107(37), 16196–16200. <https://doi.org/10.1073/pnas.1006538107>
- Ferris, S. D., Sage, R. D., Prager, E. M., Ritte, U., & Wilson, A. C. (1983). Mitochondrial DNA evolution in mice. *Genetics*, 105(3), 681–721. <https://doi.org/10.1093/genetics/105.3.681>
- Fierst, J. L., & Hansen, T. F. (2010). Genetic architecture and postzygotic reproductive isolation: Evolution of Bateson–Dobzhansky–Muller incompatibilities in a polygenic model. *Evolution*, 64(3), 675–693. <https://doi.org/10.1111/j.1558-5646.2009.00861.x>
- Finger, N., Farleigh, K., Bracken, J. T., Leaché, A. D., François, O., Yang, Z., Flouri, T., Charran, T., Jezkova, T., Williams, D. A., & Blair, C. (2022). Genome-scale data reveal deep lineage divergence and a complex demographic history in the Texas horned lizard (*Phrynosoma cornutum*) throughout the southwestern and Central United States. *Genome Biology and Evolution*, 14(1), evab260. <https://doi.org/10.1093/gbe/evab260>
- Fleishman, L. J., Leal, M., & Persons, M. H. (2009). Habitat light and dewlap color diversity in four species of Puerto Rican anoline lizards. *Journal of Comparative Physiology A*, 195(11), 1043–1060. <https://doi.org/10.1007/s00359-009-0478-8>
- Fleishman, L. J., Ogas, B., Steinberg, D., & Leal, M. (2016). Why do *Anolis* dewlaps glow? An analysis of a translucent visual signal. *Functional Ecology*, 30(3), 345–355. <https://doi.org/10.1111/1365-2435.12502>
- Garrison, E., & Marth, G. (2012). Haplotype-based variant detection from short-read sequencing. *arXiv*, 1207.3907v2.
- Gorman, G. C., & Atkins, L. (1969). The zoogeography of lesser Antillean *Anolis* lizards: An analysis based upon chromosomes and lactic dehydrogenases. *Bulletin of the Museum of Comparative Zoology*, 138(3), 53–80.
- Gorman, G. C., & Stamm, B. (1975). The *Anolis* lizards of Mona, Redonda, and La Blanquilla: Chromosomes, relationships, and natural history notes. *Journal of Herpetology*, 9(2), 197–205. <https://doi.org/10.2307/1563037>
- Gow, J. L., Peichel, C. L., & Taylor, E. B. (2006). Contrasting hybridization rates between sympatric three-spined sticklebacks highlight the fragility of reproductive barriers between evolutionarily young species. *Molecular Ecology*, 15(3), 739–752. <https://doi.org/10.1111/j.1365-294X.2006.02825.x>
- Grant, P. R., & Grant, B. R. (1994). Phenotypic and genetic effects of hybridization in Darwin's finches. *Evolution*, 48(2), 297–316. <https://doi.org/10.1111/j.1558-5646.1994.tb01313.x>
- Gunderson, A. R., & Leal, M. (2016). A conceptual framework for understanding thermal constraints on ectotherm activity with implications for predicting responses to global change. *Ecology Letters*, 19(2), 111–120. <https://doi.org/10.1111/ele.12552>
- Gunderson, A. R., Mahler, D. L., & Leal, M. (2018). Thermal niche evolution across replicated *Anolis* lizard adaptive radiations. *Proceedings of the Royal Society B: Biological Sciences*, 285(1877), 1–9. <https://doi.org/10.1098/rspb.2017.2241>
- Harrison, R. G., & Larson, E. L. (2014). Hybridization, introgression, and the nature of species boundaries. *Journal of Heredity*, 105(S1), 795–809. <https://doi.org/10.1093/jhered/esu033>
- Harrison, R. G., & Larson, E. L. (2016). Heterogeneous genome divergence, differential introgression, and the origin and structure of hybrid zones. *Molecular Ecology*, 25(11), 2454–2466. <https://doi.org/10.1111/mec.13582>
- Hedrick, P. W. (2013). Adaptive introgression in animals: Examples and comparison to new mutation and standing variation as sources of adaptive variation. *Molecular Ecology*, 22(18), 4606–4618. <https://doi.org/10.1111/mec.12415>
- Hill, G. E., Havird, J. C., Sloan, D. B., Burton, R. S., Greening, C., & Dowling, D. K. (2019). Assessing the fitness consequences of mitochondrial interactions in natural populations. *Biological Reviews*, 94(3), 1089–1104. <https://doi.org/10.1111/brv.12493>
- Hird, S., & Sullivan, J. (2009). Assessment of gene flow across a hybrid zone in red-tailed chipmunks (*Tamias ruficaudus*). *Molecular Ecology*, 18(14), 3097–3109. <https://doi.org/10.1111/j.1365-294X.2009.04196.x>
- Hohenlohe, P. A., Bassham, S., Etter, P. D., Stiffler, N., Johnson, E. A., & Cresko, W. A. (2010). Population genomics of parallel adaptation in threespine stickleback using sequenced RAD tags. *PLoS Genetics*, 6(2), e1000862. <https://doi.org/10.1371/journal.pgen.1000862>
- Huey, R. B., Losos, J. B., & Moritz, C. (2010). Are lizards toast? *Science*, 328(5980), 832–833. <https://doi.org/10.1126/science.1190374>
- Jezkova, T., Leal, M., & Rodríguez-Robles, J. A. (2013a). Genetic drift or natural selection? Hybridization and asymmetric mitochondrial introgression in two Caribbean lizards (*Anolis pulchellus* and *Anolis krugi*). *Journal of Evolutionary Biology*, 26(7), 1458–1471. <https://doi.org/10.1111/jeb.12149>
- Jezkova, T., Leal, M., & Rodríguez-Robles, J. A. (2013b). Data from: Genetic drift or natural selection? Hybridization and asymmetric mitochondrial introgression in two Caribbean lizards (*Anolis pulchellus* and *Anolis krugi*). *Dryad Dataset*. <https://doi.org/10.5061/dryad.304dn>
- Jombart, T. (2008). ADEGENET: A R package for the multivariate analysis of genetic markers. *Bioinformatics*, 24(11), 1403–1405. <https://doi.org/10.1093/bioinformatics/btn129>
- Jones, F. C., Grabherr, M. G., Chan, Y. F., Russell, P., Mauceli, E., Johnson, J., Swofford, R., Pirun, M., Zody, M. C., White, S., Birney, E., Searle, S., Schmutz, J., Grimwood, J., Dickson, M. C., Myers, R. M., Miller, C. T., Summers, B. R., Knecht, A. K., ... Kingsley, D. M. (2012). The genomic basis of adaptive evolution in threespine sticklebacks. *Nature*, 484(7392), 55–61. <https://doi.org/10.1038/nature10944>
- Jouganous, J., Long, W., Ragsdale, A. P., & Gravel, S. (2017). Inferring the joint demographic history of multiple populations: Beyond the diffusion approximation. *Genetics*, 206(3), 1549–1567. <https://doi.org/10.1534/genetics.117.200493>
- Leaché, A. D., Portik, D. M., Rivera, D., Rödel, M. O., Penner, J., Gvoždík, V., Greenbaum, E., Jongsma, G. F. M., Ofori-Boateng, C., Burger, M., Eniang, E. A., Bell, R. C., & Fujita, M. K. (2019). Exploring rain forest diversification using demographic model testing in the African foam-nest treefrog *Chiromantis rufescens*. *Journal of Biogeography*, 46(12), 2706–2721. <https://doi.org/10.1111/jbi.13716>
- Leal, M. (1999). Honest signalling during prey–predator interactions in the lizard *Anolis cristatellus*. *Animal Behaviour*, 58(3), 521–526. <https://doi.org/10.1006/anbe.1999.1181>
- Leal, M., & Losos, J. B. (2015). A naturalist's insight into the evolution of signal redundancy. *The American Naturalist*, 186(4), ii–iv. <https://doi.org/10.1086/682704>
- Li, H. (2013). *Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM*. arXiv:1303.3997v1.
- Lischer, H. E., & Excoffier, L. (2012). PGDSpider: An automated data conversion tool for connecting population genetics and genomics programs. *Bioinformatics*, 28(2), 298–299. <https://doi.org/10.1093/bioinformatics/btr642>
- Liu, K., Wang, F., Chen, W., Tu, L., Min, M. S., Bi, K., & Fu, J. (2010). Rampant historical mitochondrial genome introgression between two species of green pond frogs, *Pelophylax nigromaculatus* and *P. plancyi*. *BMC Evolutionary Biology*, 10(1), 1–14. <https://doi.org/10.1186/1471-2148-10-201>
- Llopart, A., Herrig, D., Brud, E., & Stecklein, Z. (2014). Sequential adaptive introgression of the mitochondrial genome in *Drosophila yakuba* and *Drosophila santomea*. *Molecular Ecology*, 23(5), 1124–1136. <https://doi.org/10.1111/mec.12678>
- Llopart, A., Lachaise, D., & Coyne, J. A. (2005). Multilocus analysis of introgression between two sympatric sister species of *Drosophila*: *Drosophila yakuba* and *D. santomea*. *Genetics*, 171(1), 197–210. <https://doi.org/10.1534/genetics.104.033597>

- López-Delgado, J., van Riemsdijk, I., & Arntzen, J. (2020). Tracing species replacement in marbled newts. *Authorea preprints*. <https://doi.org/10.22541/au.158488953.30271975>
- Losos, J. B. (1990). Ecomorphology, performance capability, and scaling of West Indian *Anolis* lizards: An evolutionary analysis. *Ecological Monographs*, 60(3), 369–388. <https://doi.org/10.2307/1943062>
- Losos, J. B. (2009). *Lizards in an evolutionary tree*. University of California Press.
- Losos, J. B., & Irschick, D. J. (1996). The effect of perch diameter on escape behaviour of *Anolis* lizards: Laboratory predictions and field tests. *Animal Behaviour*, 51(3), 593–602. <https://doi.org/10.1006/anbe.1996.0063>
- Losos, J. B., Jackman, T. R., Larson, A., De Queiroz, K., & Rodríguez-Schettino, L. (1998). Contingency and determinism in replicated adaptive radiations of Island lizards. *Science*, 279(5359), 2115–2118. <https://doi.org/10.1126/science.279.5359.2115>
- Losos, J. B., Warheit, K. I., & Schoener, T. W. (1997). Adaptive differentiation following experimental Island colonization in *Anolis* lizards. *Nature*, 387(6628), 70–73. <https://doi.org/10.1038/387070a0>
- Mallet, J. (2005). Hybridization as an invasion of the genome. *Trends in Ecology and Evolution*, 20(5), 229–237. <https://doi.org/10.1016/j.tree.2005.02.010>
- Martin, B. T., Douglas, M. R., Chafin, T. K., Placyk, J. S., Birkhead, R. D., Phillips, C. A., & Douglas, M. E. (2020). Contrasting signatures of introgression in North American box turtle (*Terrapene* spp.) contact zones. *Molecular Ecology*, 29(21), 4186–4202. <https://doi.org/10.1111/mec.15622>
- Martin, S. H., Davey, J. W., Salazar, C., & Jiggins, C. D. (2019). Recombination rate variation shapes barriers to introgression across butterfly genomes. *PLoS Biology*, 17(2), e2006288. <https://doi.org/10.1371/journal.pbio.2006288>
- Mašek, J., & Andersson, E. R. (2017). The developmental biology of genetic Notch disorders. *Development*, 144(10), 1743–1763. <https://doi.org/10.1242/dev.148007>
- Massey Jr, F. J. (1951). The Kolmogorov-smirnov test for goodness of fit. *Journal of the American statistical Association*, 46(253), 68–78. <https://doi.org/10.1080/01621459.1951.10500769>
- Morales, H. E., Sunnucks, P., Joseph, L., & Pavlova, A. (2017). Perpendicular axes of differentiation generated by mitochondrial introgression. *Molecular Ecology*, 26(12), 3241–3255. <https://doi.org/10.1111/mec.14114>
- Nosil, P. (2012). *Ecological speciation*. Oxford University Press.
- Pardo-Díaz, C., Salazar, C., Baxter, S. W., Merot, C., Figueiredo-Ready, W., Joron, M., McMillan, W. O., & Jiggins, C. D. (2012). Adaptive introgression across species boundaries in *Heliconius* butterflies. *PLoS Genetics*, 8(6), e1002752. <https://doi.org/10.1371/journal.pgen.1002752>
- Pardo-Díaz, C., Salazar, C., & Jiggins, C. D. (2015). Towards the identification of the loci of adaptive evolution. *Methods in Ecology and Evolution*, 6(4), 445–464. <https://doi.org/10.1111/2041-210X.12324>
- Payseur, B. A. (2010). Using differential introgression in hybrid zones to identify genomic regions involved in speciation. *Molecular Ecology Resources*, 10(5), 806–820. <https://doi.org/10.1111/j.1755-0998.2010.02883.x>
- Peterson, B. K., Weber, J. N., Kay, E. H., Fisher, H. S., & Hoekstra, H. E. (2012). Double digest RADseq: An inexpensive method for de novo SNP discovery and genotyping in model and non-model species. *PLoS One*, 7(5), e37135. <https://doi.org/10.1371/journal.pone.0037135>
- Petr, M., Pääbo, S., Kelso, J., & Vernet, B. (2019). Limits of long-term selection against Neandertal introgression. *Proceedings of the National Academy of Sciences of the United States of America*, 116(5), 1639–1644. <https://doi.org/10.1073/pnas.1814338116>
- Portik, D. M., Leaché, A. D., Rivera, D., Barej, M. F., Burger, M., Hirschfeld, M., Rödel, M. O., Blackburn, D. C., & Fujita, M. K. (2017). Evaluating mechanisms of diversification in a Guineo-Congolian tropical forest frog using demographic model selection. *Molecular Ecology*, 26(19), 5245–5263. <https://doi.org/10.1111/mec.14266>
- Prentki, M., & Madiraju, S. M. (2008). Glycerolipid metabolism and signaling in health and disease. *Endocrine Reviews*, 29(6), 647–676. <https://doi.org/10.1210/er.2008-0007>
- Puritz, J. B., Hollenbeck, C. M., & Gold, J. R. (2014). dDocent: A RADseq, variant-calling pipeline designed for population genomics of non-model organisms. *PeerJ*, 2, e431. <https://doi.org/10.7717/peerj.431>
- Puritz, J. B., Matz, M. V., Toonen, R. J., Weber, J. N., Bolnick, D. I., & Bird, C. E. (2014). Demystifying the RAD fad. *Molecular Ecology*, 23(24), 5937–5942. <https://doi.org/10.1111/mec.12965>
- R Core Team. (2020). *R: A language and environment for statistical computing* (4.0.2). R Foundation for Statistical Computing. <https://www.r-project.org/>
- Rand, D. M., Fry, A., & Sheldahl, L. (2006). Nuclear-mitochondrial epistasis and *drosophila* aging: Introgression of *Drosophila simulans* mtDNA modifies longevity in *D. melanogaster* nuclear backgrounds. *Genetics*, 172(1), 329–341. <https://doi.org/10.1534/genetics.105.046698>
- Rieseberg, L. H. (2011). Adaptive introgression: The seeds of resistance. *Current Biology*, 21(15), R581–R583. <https://doi.org/10.1016/j.cub.2011.06.038>
- Rieseberg, L. H., Sinervo, B., Linder, C. R., Ungerer, M. C., & Arias, D. M. (1996). Role of gene interactions in hybrid speciation: Evidence from ancient and experimental hybrids. *Science*, 272(5262), 741–745. <https://doi.org/10.1126/science.272.5262.741>
- Rivero, J. A. (1998). *Amphibians and reptiles of Puerto Rico*. University of Puerto Rico Press.
- Rodríguez-Robles, J. A., Jezkova, T., & Leal, M. (2010). Climatic stability and genetic divergence in the tropical insular lizard *Anolis krugi*, the Puerto Rican “Lagartijo Jardinero de la Montaña”. *Molecular Ecology*, 19(9), 1860–1876. <https://doi.org/10.1111/j.1365-294X.2010.04616.x>
- Schliep, K. P. (2011). phangorn: Phylogenetic analysis in R. *Bioinformatics*, 27(4), 592–593. <https://doi.org/10.1093/bioinformatics/btq706>
- Schluter, D. (2000). *The ecology of adaptive radiation*. Oxford University Press.
- Schumer, M., Xu, C., Powell, D. L., Durvasula, A., Skov, L., Holland, C., Blazier, J. C., Sankararaman, S., Andolfatto, P., Rosenthal, G. G., & Przeworski, M. (2018). Natural selection interacts with recombination to shape the evolution of hybrid genomes. *Science*, 360(6389), 656–660. <https://doi.org/10.1126/science.aar3684>
- Schwartz, A., & Henderson, R. W. (1991). *Amphibians and reptiles of the West Indies: Descriptions, distributions, and natural history*. University Press of Florida.
- Seehausen, O. (2004). Hybridization and adaptive radiation. *Trends in Ecology and Evolution*, 19(4), 198–207. <https://doi.org/10.1016/j.tree.2004.01.003>
- Seehausen, O. (2006). Conservation: Losing biodiversity by reverse speciation. *Current Biology*, 16(9), 334–337. <https://doi.org/10.1016/j.cub.2006.03.077>
- Sharma, L. K., Lu, J., & Bai, Y. (2009). Mitochondrial respiratory complex I: Structure, function and implication in human diseases. *Current Medicinal Chemistry*, 16(10), 1266–1277. <https://doi.org/10.2174/092986709787846578>
- Shi, X., & Wang, R. (2017). Glioma cell fate decisions mediated by Dll1-Jag1-Fringe in Notch1 signaling pathway. *BMC Systems Biology*, 11(4), 75–84. <https://doi.org/10.1186/s12918-017-0457-6>
- Sloan, D. B., Havird, J. C., & Sharbrough, J. (2017). The on-again, off-again relationship between mitochondrial genomes and species boundaries. *Molecular Ecology*, 26(8), 2212–2236. <https://doi.org/10.1111/mec.13959>
- Stroud, D. A., Surgenor, E. E., Formosa, L. E., Reljic, B., Frazier, A. E., Dibley, M. G., Osellame, L. D., Stait, T., Beilharz, T. H., Thorburn,

- D. R., Salim, A., & Ryan, M. T. (2016). Accessory subunits are integral for assembly and function of human mitochondrial complex I. *Nature*, 538(7623), 123–126. <https://doi.org/10.1038/nature19754>
- Taylor, E. B., Boughman, J. W., Groenenboom, M., Sniatynski, M., Schluter, D., & Gow, J. L. (2006). Speciation in reverse: Morphological and genetic evidence of the collapse of a three-spined stickleback (*Gasterosteus aculeatus*) species pair. *Molecular Ecology*, 15(2), 343–355. <https://doi.org/10.1111/j.1365-294X.2005.02794.x>
- Tegelström, H. (1987). Transfer of mitochondrial DNA from the northern red-backed vole (*Clethrionomys rutilus*) to the bank vole (*C. glareolus*). *Journal of Molecular Evolution*, 24(3), 218–227. <https://doi.org/10.1007/BF02111235>
- Tickle, C., & Towers, M. (2017). Sonic hedgehog signaling in limb development. *Frontiers in Cell and Developmental Biology*, 5, 14. <https://doi.org/10.3389/fcell.2017.00014>
- Tigano, A., Shultz, A. J., Edwards, S. V., Robertson, G. J., & Friesen, V. L. (2017). Outlier analyses to test for local adaptation to breeding grounds in a migratory arctic seabird. *Ecology and Evolution*, 7(7), 2370–2381. <https://doi.org/10.1002/ece3.2819>
- Toews, D. P. L., & Brelsford, A. (2012). The biogeography of mitochondrial and nuclear discordance in animals. *Molecular Ecology*, 21(16), 3907–3930. <https://doi.org/10.1111/j.1365-294X.2012.05664.x>
- Tollis, M., & Boissinot, S. (2014). Genetic variation in the green anole lizard (*Anolis carolinensis*) reveals Island refugia and a fragmented Florida during the quaternary. *Genetica*, 142(1), 59–72. <https://doi.org/10.1007/s10709-013-9754-1>
- Verhoeven, K. J., Simonsen, K. L., & McIntyre, L. M. (2005). Implementing false discovery rate control: Increasing your power. *Oikos*, 108(3), 643–647. <https://doi.org/10.1111/j.0030-1299.2005.13727.x>
- Walsh, J., Shriver, W. G., Olsen, B. J., & Kovach, A. I. (2016). Differential introgression and the maintenance of species boundaries in an advanced generation avian hybrid zone. *BMC Evolutionary Biology*, 16(1), 1–18. <https://doi.org/10.1186/s12862-016-0635-y>
- Warheit, K. I., Forman, J. D., Losos, J. B., & Miles, D. B. (1999). Morphological diversification and adaptive radiation: A comparison of two diverse lizard clades. *Evolution*, 53(4), 1226–1234. <https://doi.org/10.1111/j.1558-5646.1999.tb04535.x>
- Whitney, K. D., Randell, R. A., & Rieseberg, L. H. (2006). Adaptive introgression of abiotic tolerance traits in the sunflower *Helianthus annuus*. *The American Naturalist*, 167(6), 230–239. <https://doi.org/10.1111/j.1469-8137.2010.03234.x>
- Williams, E. E. (1983). Ecomorphs, faunas, Island size, and diverse end points in Island radiations of *Anolis*. In R. B. Huey, E. R. Pianka, & T. W. Schoener (Eds.), *Lizard ecology: Studies of a model organism* (pp. 326–370). Harvard University Press.
- Wolff, J. N., Ladoukakis, E. D., Enriquez, J. A., & Dowling, D. K. (2014). Mitonuclear interactions: Evolutionary consequences over multiple biological scales. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 369(1646), 20130443. <https://doi.org/10.1098/rstb.2013.0443>
- Yan, L., Jin, W., Zhao, Q., Cui, X., Shi, T., Xu, Y., Li, F., Jin, W., Zhang, Z., Zhang, Z., Tang, Q.-Q., & Pan, D. (2021). PWWP2B fine-tunes adipose thermogenesis by stabilizing HDACs in a NuRD subcomplex. *Advanced Science*, 8(16), 2102060. <https://doi.org/10.1002/adv.202102060>

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Farleigh, K., Ascanio, A., Farleigh, M. E., Schield, D. R., Card, D. C., Leal, M., Castoe, T. A., Jezkova, T., & Rodríguez-Robles, J. A. (2023). Signals of differential introgression in the genome of natural hybrids of Caribbean anoles. *Molecular Ecology*, 32, 6000–6017. <https://doi.org/10.1111/mec.17170>